

CERTIFICATE

Name: Vinita Kumawat

Class: BSC I year Sem II

Roll No.: 131

Exam No.:

Institution Vivek P.G. College, Kalwar

This is certified to be the bonafide work of the student in the Botany
Semester II Laboratory during the academic
year 2023 /2024

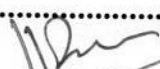
No of practicals certified 10 out of 12 in the
subject of Botany



Examiner's Signature

Date : 12 May 2024

.....
Teacher in-charge

.....

Principal
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KALWAR, JAIPUR-303706
institution Rubber stamp

Vinita Kumawat


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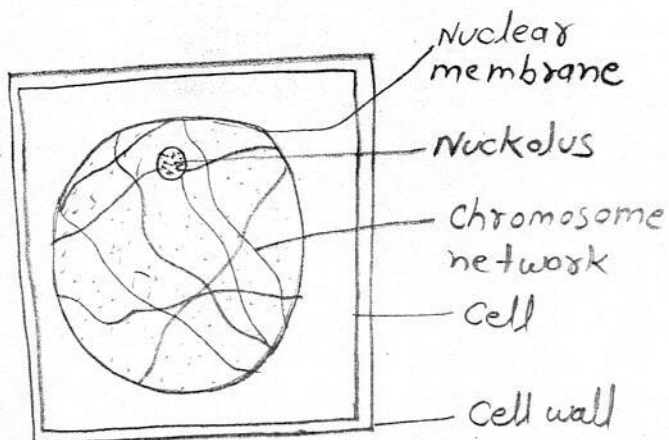
S. No.	Name of Experiment	Page No.	Date of Experiment	Date of Submission	Remarks
1.	Study of Meiotic Division Meiosis - I	1-2	13/03/24	14/03/24	Submitted 14/03/24
2.	Salivary Gland Polytene Chromosome	3	14/03/24	15/03/24	Submitted 15/03/24
3.	Structure of Cell Onion Cell	4	16/03/24	18/03/24	Submitted 18/03/24
4.	Spirogyra	5	19/03/24	20/03/24	Submitted 20/03/24
5.	Study Electronmicrograph of Eucaryotic cell organelles	6	22/03/24	23/03/24	Submitted 23/03/24
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(2)		8	22/03/24	23/03/24	Submitted 23/03/24
(3)	Nucleus माइक्रोकेंद्रिका	9	22/03/24	23/03/24	Submitted 23/03/24
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(9)	Golgi bodies	15	30/03/24	01/04/24	Submitted 01/04/24
(10)	Endoplasmic Reticulum	16	30/03/24	01/04/24	Submitted 01/04/24
<u>Genetics</u>		17-25			
6.	Monohybrid cross	17	03/04/24	05/04/24	Submitted 05/04/24
7.	Dihybrid cross	18	7/04/24	12/04/24	Submitted 12/04/24
8.	Mendel's Rules	19	15/04/24	16/04/24	Submitted 16/04/24
9.	Test cross & Back cross	20	18/04/24	19/04/24	Submitted 19/04/24

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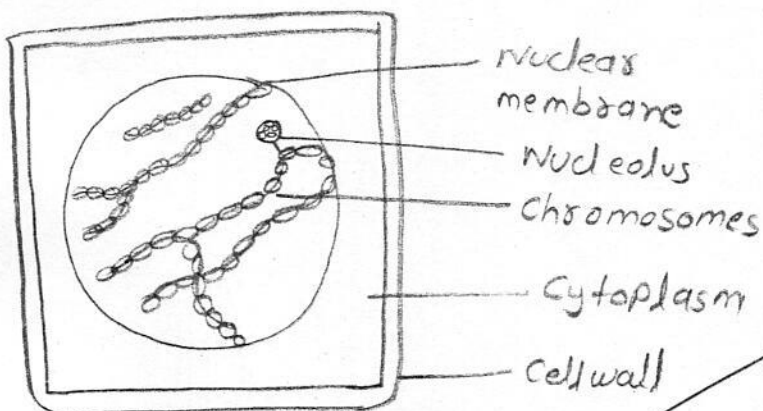
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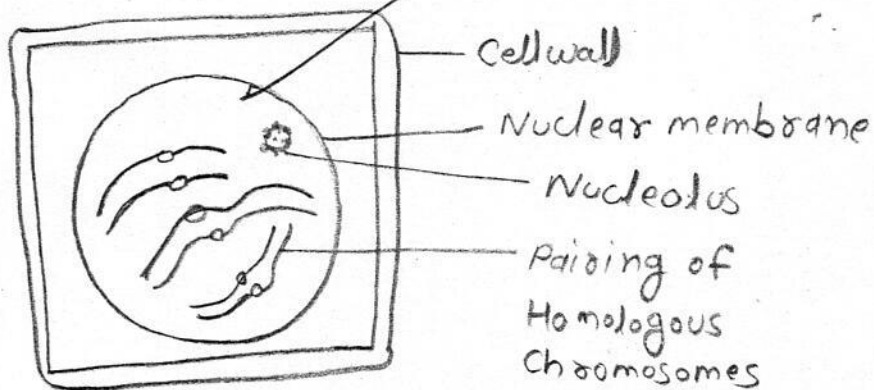

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PROPHASE



(A) Leptotene



(B) Zygotene

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Study of Meiotic Division

Meiosis - I

प्रेक्षण

1. प्रथम अर्द्धसूत्री विभाजन

Prophase - I

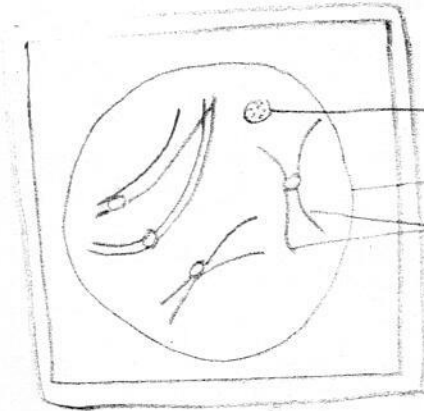
यह अर्द्धसूत्री विभाजन की प्रथम व सबसे लंबी प्रावस्था होती है। यह पांच उप-प्रावस्थाओं में जैसे लेप्टोटीन, जायगोटीन, पैकीटीन, डिप्लोटीन व डाइकार्नेसिस में विभाजित रहती है जो आगे वर्णित है।

पूर्वावस्था की विभिन्न अवस्थायें

Stages	Process
1. Leptoteme	आरस्रष्ट व प्रोटीन के निर्माण द्वारा केन्द्रक के आकार में वृद्धि, गुणसूत्रों का कुण्डलन द्वारा छोटे व सघट्ट होना।
2. Zygoteme	समजात गुणसूत्रों का युग्मन, सूत्रियुग्मन, दोनों अर्द्धगुणसूत्र द्वारा सिनेमिडिनिमल संकुल का निर्माण।
3. Pachytene	क्रॉसिंग ओवर, विजातीय गुणसूत्र में जीन विनिमय।
4. Diplotene	समजात गुणसूत्रों का पुनिकर्षण बल द्वारा अलग-2 होना कायज्मेटा का दिखाई देना कायज्मेटा की गुणसूत्रों की भुजायें का 180° कोण पर घुम जाना। x जैसा।
5. Diakinesis	कायज्मेटा का पूर्ण रूप से उपांतीकरण व क्रोमोसोम स्म का संघनन।

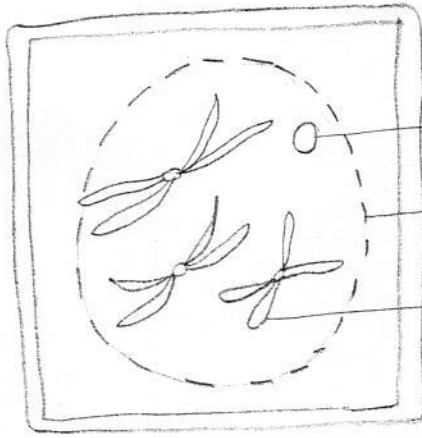
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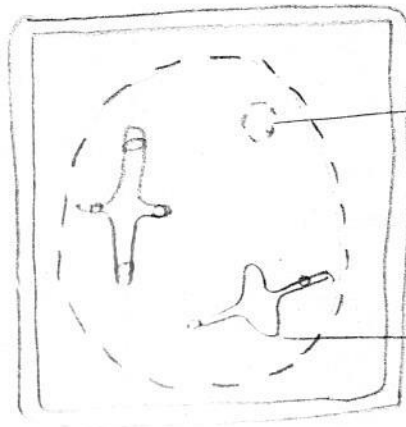
nucleolus
 Nuclear membrane
 Pair of sister chromatids

(c) Pachytene



Nucleolus and Nuclear me.
 disappearing
 Homologous chromosomes

(d) Diplotene



Nucleolus and Nuclear
 Homologous chromosomes

(e) Diakinesis

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Leptoteme :- 1. इसमें क्रोमोसोम लंबे, पतले, अर्न्तगुंथे धागे समान दिखाई देते हैं।

2. क्रोमोसोम की उपास्थिति के कारण यह मालाकत आभासित होते हैं।
3. इस अवस्था में क्रोमोसोम द्विगुणित अवस्था प्रदर्शित करते हैं।

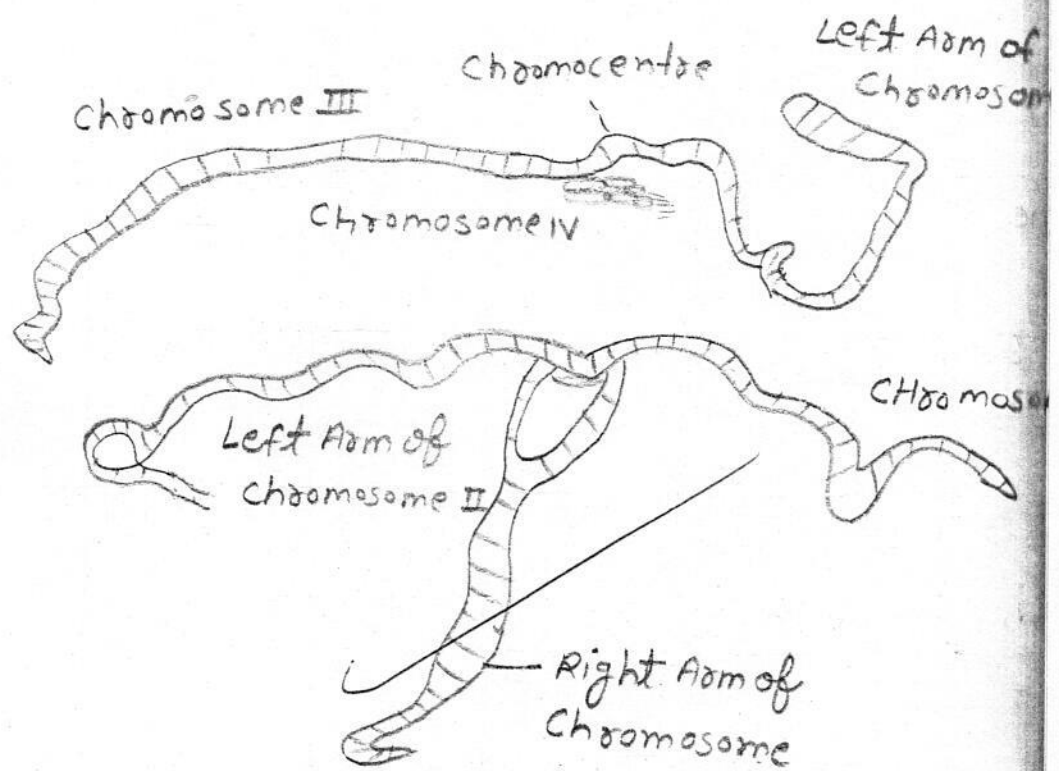
Zygoteme :- 1. जोड़ा बनाने की यह प्रक्रिया सिनेप्सिस कहलाती है।
2. यह सिनेप्सिस बहुत सटीक बनते हैं।
3. धागे - 2 क्रोमोसोम छोटे व मोटे होते जाते हैं।

Pachytene :- 1. क्रोमोसोम के दो धागे क्रोमैटिड कहलाते हैं।
2. एक क्रोमोसोम से क्रोमैटिड भागिनी क्रोमैटिड कहलाते हैं।
3. इस उप अवस्था में काथेस्मेटा दिखाई देते हैं जो X आकार के होते हैं।

Diploteme :- 1. क्रोमैटिड के धागे एक दूसरे से पृथक् होते हैं।
2. इस दिखाई देते हैं।
3. प्रत्येक सिनेप्सिस युक्त जोड़े में चार धागे उप. होते हैं।
4. इस समय क्रोमोसोम मोटे व छोटे होते हैं।

Diakinesis :- 1. टेट्राड संकुचित होकर केन्द्रक में पृथक् रूप से दिखाई देने लगते हैं।
2. प्रत्येक क्रोसोम क्रोमोसोम और अधिक सरपट्ट हो जाते हैं।
3. केन्द्रिका व केन्द्रक झिल्ली दिखाई देने लगती है।

~~Signature~~
14/03/24



पॉलीटीन गुणसूत्र की संरचना

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Salivary Gland Polytene Chromosome

Practical Exercise :- To study Salivary gland Chromosome

Requirement :- काइरोनोमस लार्वा अथवा लार्वाणि गुणसूत्र का चार्ट, नोट बुक, पेन, नीडल

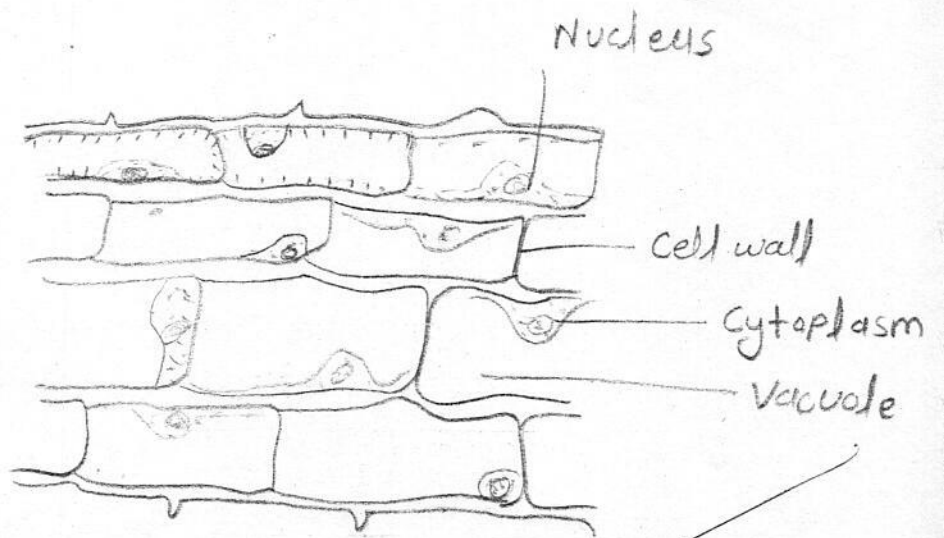
Structure of Salivary gland Chromosomes :-

1. यह डी.जी. बैलबियानी द्वारा 1881 में रिपोर्ट किये गये थे।
2. यह विशिष्ट गुणसूत्र आकार में काफी बड़े होते हैं।
3. यह सामान्य गुणसूत्रों से 200 गुणा तक बड़े हो सकते हैं।
4. अतिरिक्त गुणसूत्रों पर गहरे क्रोमैटिक व हल्के क्रोमैटिक रंग की एकान्तरित अनुप्रस्थ पट्टियां पाई जाती हैं।
5. पट्टी क्षेत्र में क्रोमोसैन्टर उप-होता है। क्रोमैनिमेटा के पार्श्व में लूप बन जाने के कारण पट्टी क्षेत्र का व्यास बढ़ जाता है।
6. यह पार्श्वीय क्षेत्र फूला होता है। तथा यह बैलबियानी तन्त्र कहलाते हैं।
7. यह सक्रिय RNA संश्लेषण के विस्थल हैं जो अनुलेखन विस्थलों को इंगित करते हैं।
8. पॉलीटीन गुणसूत्र चिर प्रोफेज अवस्था में पाये जाते हैं।


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15/3/24

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पत्र की पत्रियों की कोशिका संरचना


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Structure of Cell

Onion Cell

Practical Exercise :- Study of Cell's in onion

Requirements :- प्याज के कंद स्लाइड, कवर स्लिप, वॉच ग्लास, आसुत जल, सैफ्रेनिन, ग्लिसरीन।

Cell :- पादप काय की सबसे छोटी संरचनात्मक तथा कार्यात्मक इकाई कोशिका कहलाती है। पादप कोशिका में शक्ति उपस्थित होती है।

Procedure :- प्याज कंद माँसल इनकी निचली सतह की बाह्य त्वचा हटा कर जल में रखें।

Cell Struc. in onion peel :- 1. पर्ण में यूकेरियोटिक कोशिका संगठन उपस्थित है।
 2. कोशिका भित्ति के भीतर प्लाज्मा उपस्थित है।
 3. प्लाज्मा झिल्ली के भीतर प्रोटोप्लाज्म उप. है। जिससे सुस्पष्ट केंद्रक पाया जाता है।
 4. प्रोटोप्लाज्म में कई डिस्क समान अवर्णी लवक उप. है।
 5. रिक्तिकायें पायी जाती हैं जिनमें कोशिका रस भरा रहता है।

Inference :- प्याज की कोशिकायें संगठित हैं। कोशिकांग कोशिका भित्ति एवं प्लाज्मा झिल्ली सुरक्षित हैं। सुस्पष्ट केंद्रक व हरितलवक उप. है। अतः यह यूकेरियोटिक Cell है।

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18/3/24

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Spirogyra

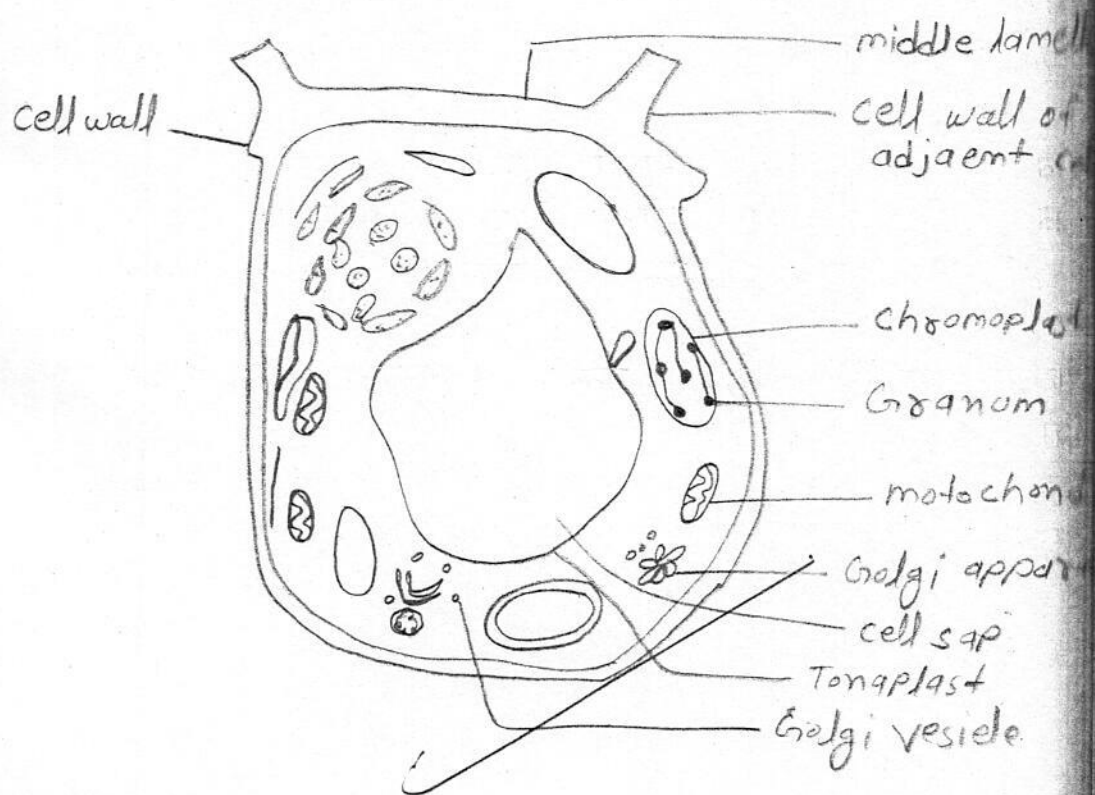
Practical Exercise :- Study of cells of algae Spirogyra

Requirements :- स्पाइरोगैरा पाकप, स्लाइड, कवर स्लिप, वाच ग्लास
आसुत जल, शै फ्रेमिन, ग्लिसरीन

Cell Structure of Spirogyra :-

1. स्पाइरोगैरा की कोशिका में यूकेरियोटिक कोशिका संगठन पाया जाता है।
2. कोशिका आयताकार होती है।
3. कोशिका के चारों ओर दृढ़ कोशिका भित्ति पायी जाती है व इसके बाहर श्लैष्मा का आवरण उपस्थित होता है।
4. कोशिका भित्ति के भीतर प्लाज्मा कला पायी जाती है।

Atul
20/3/24



पादप कोशिका की परासंरचना

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Study of Electronmicrograph of Eucaryotic Cell Organelles

Practical Exercise :- Study of Various organelles of Eucaryotic Cell using electronmicrograph

Requirements :- यूकेरियोटिक कोशिका जीव का इलेक्ट्रॉन माइक्रोग्राफ

● Structure of Eucaryotic Cell :- यूकेरियोटिक कोशिका जटिल होती है। माइटोकॉन्ड्रिया की संख्या एक कोशिका में 1700 तक हो सकती है। परऑक्सीसोम 400 तथा लाइसोसोम 300 तक हो सकते हैं। एक यूकेरियोटिक कोशिका में निम्नलिखित कोशिकांग उप. हो सकते हैं।

1. कोशिका भित्ति
2. प्लाज्मा कला
3. केन्द्रक
4. माइटोकॉन्ड्रिया
5. हरित लवक
6. राइबोसोम
7. परऑक्सीसोम
8. लाइसोसोम
9. गॉल्जी सामिश्र
10. अन्तः पदव्यी जालिका

कौशिका द्रव्य
यूकेरियोटिक कोशिका की परासंरचना जन्तु एवं पादप की अलग-2 चित्रों में है।

[Signature]
23/03/24

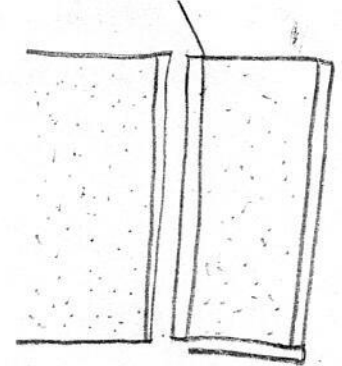
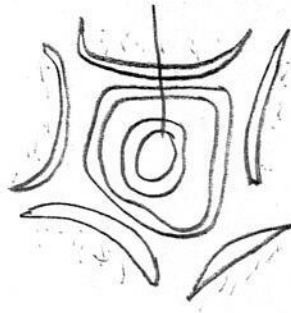
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Central lumen

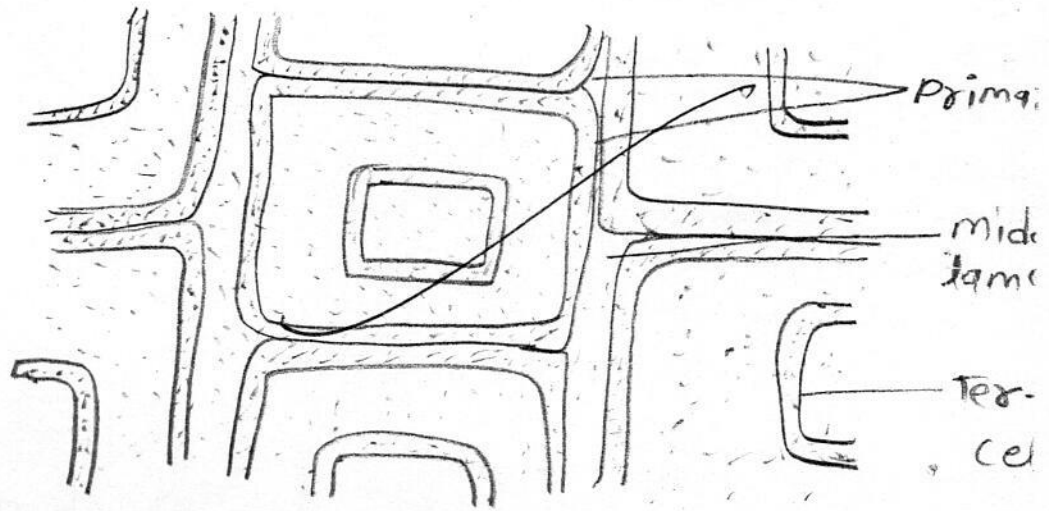
middle lamella



(A)

(B)

कोशिका भित्ति की संरचना



कोशिका का भित्ति की संरचना प्राथमिक द्वितीय
तृतीयक भित्ति

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1. Cell wall

यह पादपों का बाहरी आवरण है जो जन्तु कोशिका में अनुपस्थित होती है। यह कठोर, बाहरी निर्जीव होता है जो कोशिका की बाहरी वातावरण से रक्षा करती है।

कोशिका भित्ति की संरचना:- यह निम्न भागों से निर्मित है-
 प्राथमिक कोशिका, द्वितीयक कोशिका
 तृतीयक कोशिका व मध्यपरतिका।

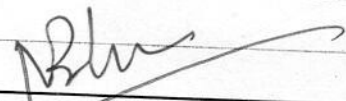
प्राथमिक कोशिका भित्ति:- यह सबसे पहले बनती जो मध्य के दोनों ओर उपस्थित रहती है। यह पतली व चिकनी होती है।

द्वितीयक भित्ति:- यह प्राथमिक भित्ति पर लिगनिन आदि पदार्थों के निक्षेपन से बनती है, कोशिका की परिपक्व अवस्था में यह मोटी, स्पष्ट व 9-10 स्थूल होती है।

मध्यपरतिका:- यह दो कोशिकाओं की कोशिका भित्ति के मध्य उपस्थित रहती है। यह कैल्सीयम पेंटेट से बनी अक्रिस्टलीय पर्त होती है।

तृतीयक कोशिका भित्ति:- यह भित्ति जायलॉन से बनी होती है जो प्रत्येक कोशिका में नहीं पायी जाती है।

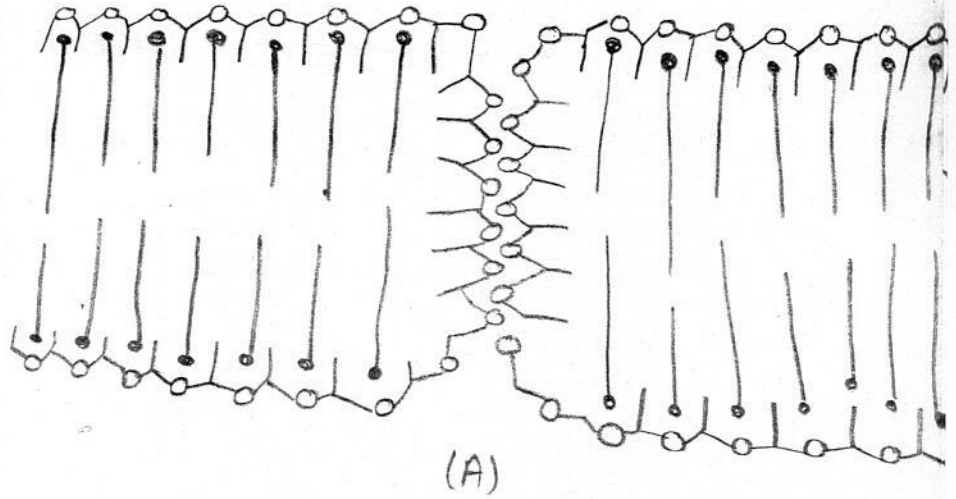
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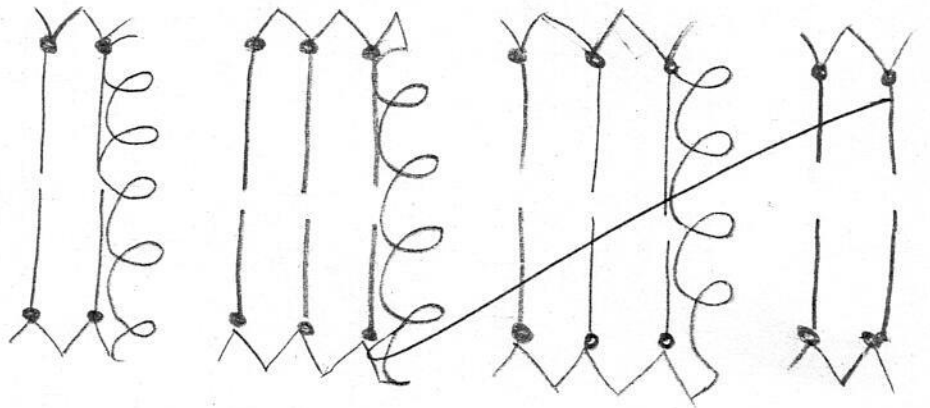
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Pg



शार्वटसन की स्क्वार्ड कला सिल्ली मॉडल

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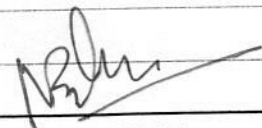
2. प्लाज्मा झिल्ली

यह चयनात्मक परागम्य झिल्ली पादप व जन्तु दोनों प्रकार की कोशिका में पायी जाती है, यह फॉस्फोलिपिड, प्रोटीन, कीलेस्ट्रॉल से बनी होती है।

रॉबर्टसन युनिट मेम्ब्रेन मॉडल :-

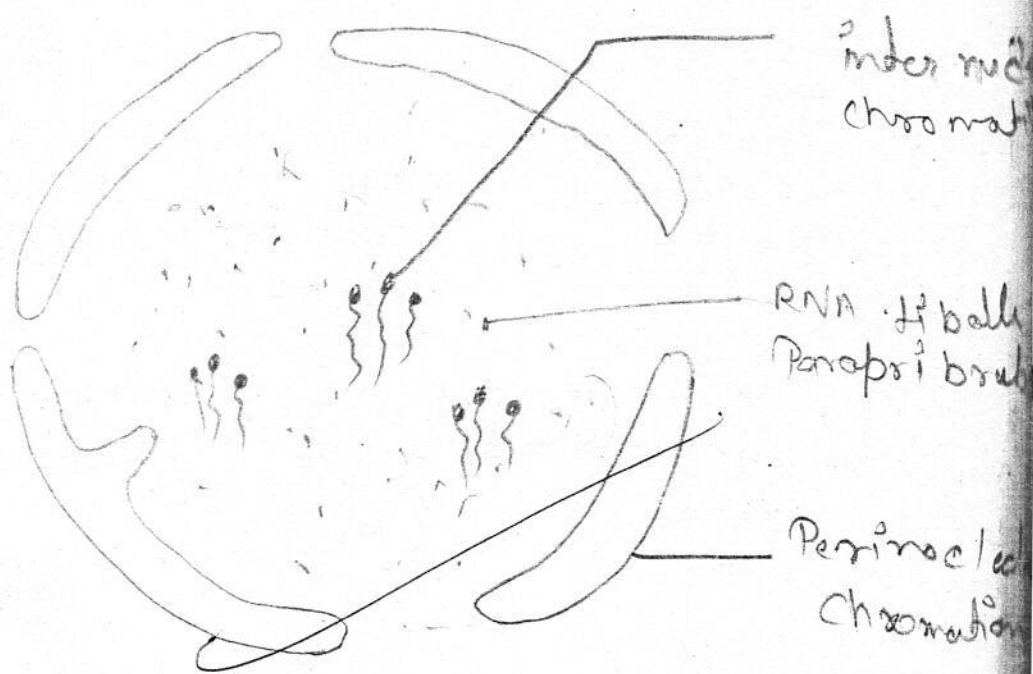
1. रॉबर्टसन ने इलेक्ट्रान माइक्रोस्कोप से अध्ययन करके प्लाज्मा झिल्ली की त्रिस्तरीय संरचना रिपोर्ट की।
2. दो बाहरी सतह $20-25 \text{ \AA}$ की होती है जो सघन ऑस्मोफिलिक होती है। यह प्रोटीन से बनी होती है।
3. मध्य पर्त हल्के रंग की $30-35 \text{ \AA}$ होती है। यह मोटी ऑस्मोफिलिक सतह होती है। यह लिपिड सतह दो प्रोटीन पर्तों के बीच उप-होती है।
4. रॉबर्टसन ने इसे स्कार्ज झिल्ली मॉडल नाम दिया।


23/03/24



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गणित विभाग

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3. केन्द्रक (Nucleus)

यह दोहरी झिल्ली से घिरा रहता है जिसमें छिद्र उप- होते हैं। इसमें केन्द्रक व क्रोमोसोम पाये जाते हैं। केन्द्रक कोशिका की सभी क्रियाओं को नियंत्रित करती है। तथा जन्म के लिये इसमें विभिन्न प्रकार के कोशिका विभाजन होते हैं।

1. केन्द्रक की संख्या :- यह सभी कोशिकाओं में पाया जाता है जबकि इसकी संख्या सब से अलग होती है।

2. केन्द्रक का आमाप :- इसका आमाप 0.5μ से 1.5μ होता है।

3. केन्द्रक का आकार :- यह कोशिका के आंतरिक वातावरण तथा क्रियात्मकता पर निर्भर करता है। यह गोलाकार, अंडाकार अथवा अन्य कई आकारों का हो सकता है। केन्द्रक का आकार इसकी अवस्था पर निर्भर करता है।

4. केन्द्रिका :- यह केन्द्रक में पायी जाने वाली एक गोलाकार संरचना है। जो प्रोटीन संश्लेषण के लिये स्थल के रूप में काम करता है।

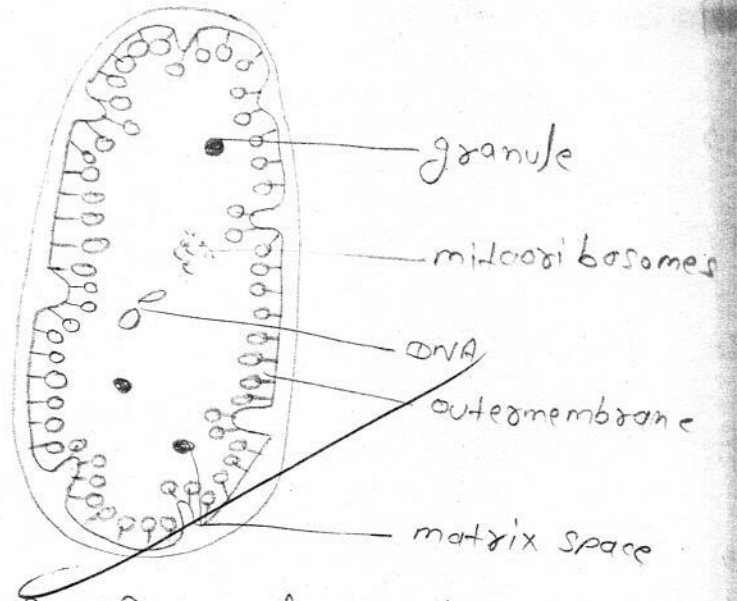
5. इसमें झिल्ली अनुपस्थित होती है। इसको 1978 में फॉन्टाना ने रिपोर्ट किया था।

~~Student~~
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चित्र - मसृटी कोशिका की परास्वरचना

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4. माइटोकॉन्ड्रिया

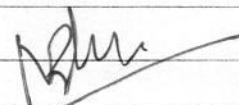
यह कोशिकाद्रव्य में उपस्थित कणिकामय सदृश्य अंगक है जो पदार्थ की ऊर्जा स्थितिज ऊर्जा को जैविक ऊर्जा में बदलती है। अतः यह कोशिका के ऊर्जा घर कहलाते हैं।

इसकी आंतरिक झिल्ली में कई बलय पाये जाते हैं। इसमें ग्लूकोस डीएनए, 10% फॉस्फेट कण, 70% प्रोटीन, 20% लिपिड तथा आर एन ए पाया जाता है। इसके मैट्रिक्स में होने वाली जैव संश्लेषण अभिक्रियाएँ हैं।

का काफी जटिल होते हैं व निम्न हैं-
 (i) शीर्ष (ii) वृत्त (iii) F_0 उपइकाई

स्त्रीगोन्ट्री कण आंतरिक सतह की m- सतह पर पाये जाते हैं जो ऑक्सीसोम या F_1 कण कहलाते हैं।

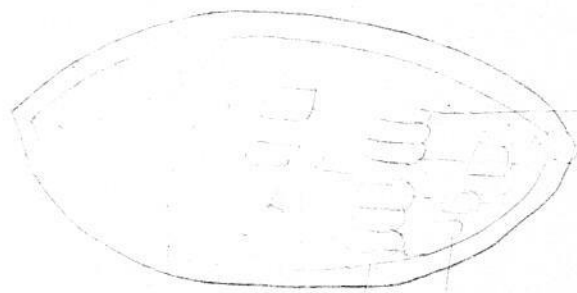
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membrane
- Plastid
- RNA

stroma

Grana lamellae

amorphous
droplet

~~चित्र - क्लोरोप्लास्ट का संरचना~~

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5. हरित लवक (Chloroplast)

हरित शोथिका पादप कोशिकाओं में हरित लवक पाया जाता है जो प्रकाश संश्लेषण में सहायता करता है।

1. हरित लवक - यह विशिष्ट कोशिकांग होते हैं जो जीवद्रव्य में बिखरे पाये जाते हैं। यह जंतु बैक्टीरिया कवक में अनुपस्थित होते हैं।

2. कुछ रंगक जैसे एन्थेसामनिन प्लास्टिड में उप. नहीं होती हैं बल्कि जीवद्रव्य में घुलित अवस्था में पाये जाते हैं।

3. हरित लवक अनेक आकार के हो सकते हैं जैसे गोल, अंडाकार, रिबन समान डिस्क समान, जालिकावत् ताराकार आदि।

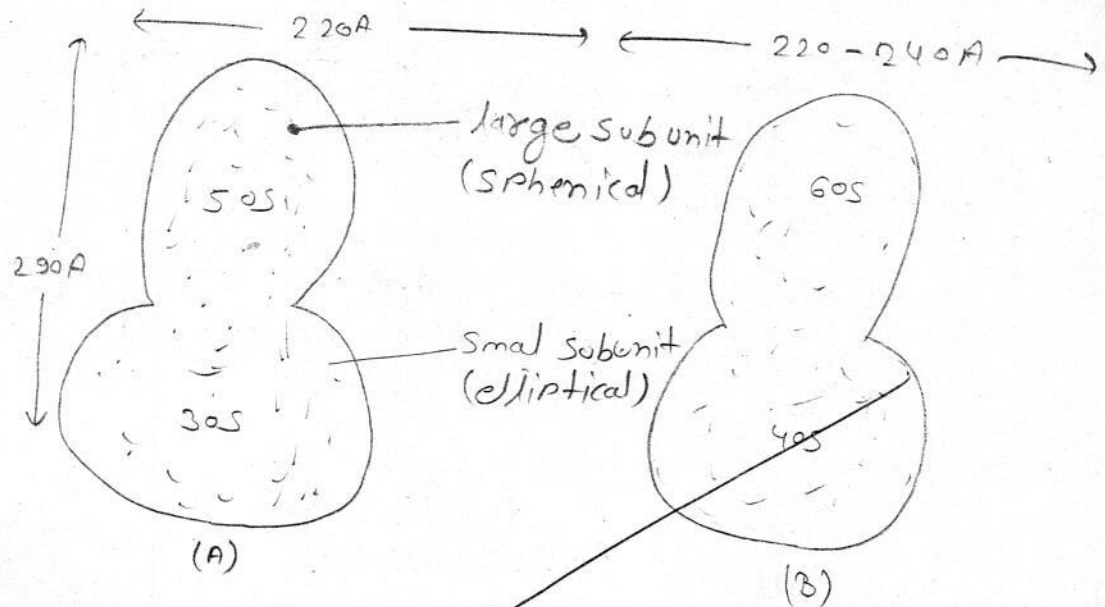
4. क्लोरोप्लास्ट का आकार इसके आकार पर निर्भर करता है सामान्यतः यह $5-6 \mu$ व्यास में $1-3 \mu$ मोटाई का होता है।

5. प्रकाश संश्लेषण की प्रकाशिक क्रिया श्रेणी में होती है।


6. क्लोरोप्लास्ट में थायलाकोइड चट्टे समान एकत्रित होकर श्रेणी का निर्माण करते हैं।

7. इसमें DNA पाया जाता है।

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चित्र - राइबोसोम

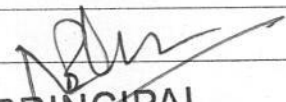

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6. राइबोसोम (Ribosome)

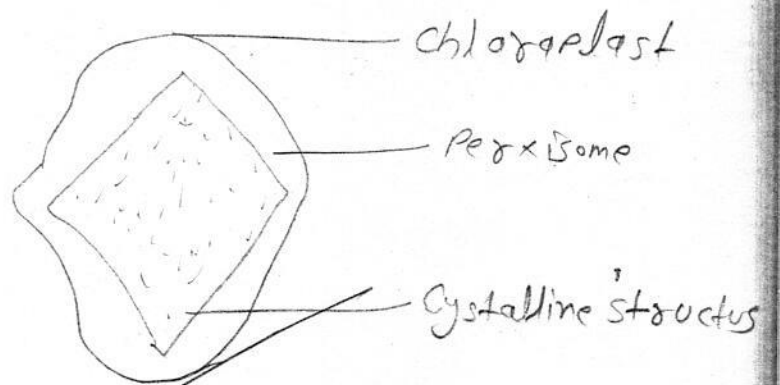
1. प्रोकैरियोटिक कोशिका का यह सबसे छोटा कोशिकांग है।
जिसमें बड़ी व छोटी उप इकाइयां होती हैं।

2. यह प्रोटीन व आरएनए से निर्मित होती है।


3. राइबोसोम प्रोटीन संश्लेषण के स्थल हैं।


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चित्र - परऑक्सीसोम


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7. परऑक्सीसोम (Peroxisomes)

1. यह गोलाकार एक कला तथा कैटलैज एन्जाइम युक्त होता है।

2. इनका प्रमुख कार्य हाइड्रोजन परऑक्साइड का अपघटन करना है।

3. पादपों में ग्लायोकजल्लेट चक्र परऑक्सीसोम में सम्पन्न होता है।

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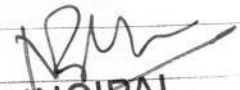
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7. परऑक्सीसोम (Peroxisomes)

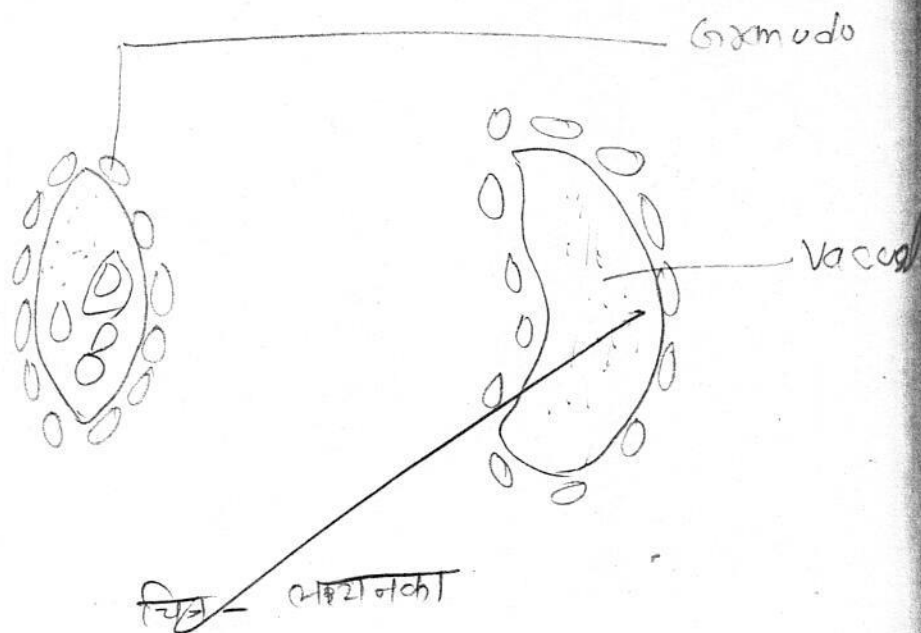
1. यह गोलाकार एक कला तथा कैटेलेज एन्जाइम युक्त होता है।
2. इनका प्रमुख कार्य हाइड्रोजन परऑक्साइड का अपघटन करना है।
3. पादपों में ग्लायोकैलेट चक्र परऑक्सीसोम में सम्पन्न होता है।


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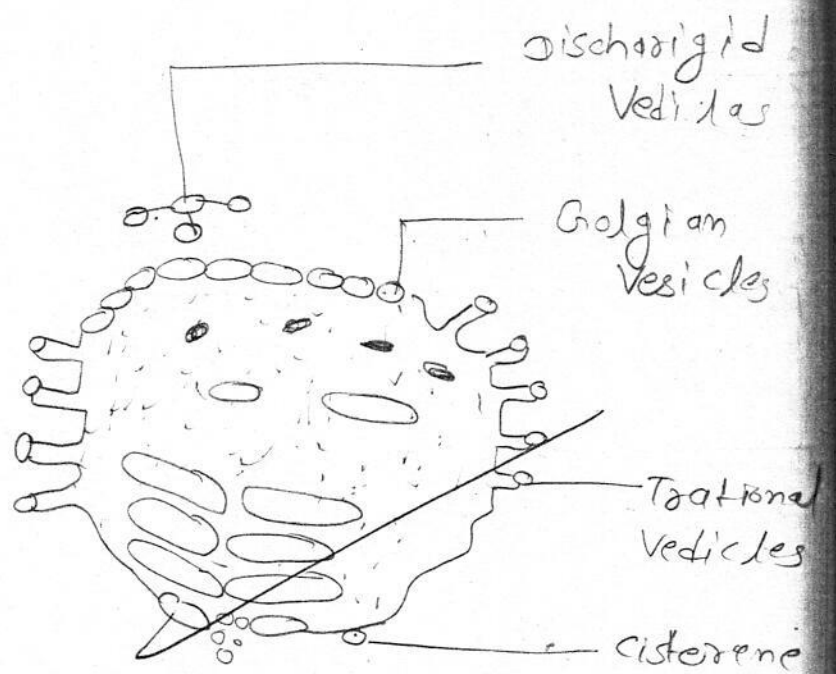

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चित्र :- गॉल्जी अण्डिका

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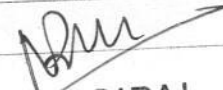
9. गॉल्जी सम्मिश्र (Golgi bodies)

1. यह प्लैट समान अनेक केन्द्रीय चपटी थैलेनुमा लुम्बी व नालिकाकार एक के ऊपर एक व्यवस्थित दूसरी का आशय निर्मित करती है।

2. इनका कार्य आन्तरिक विधि व स्थानान्तरण सिस्टम ER से प्रोटीन, गॉल्जी से दूसरे अंगों की ओर जाने का संसंचरण तथा मीसोजोम का निर्माण करना है।

3. यह ग्लाइकोप्रोटीन तथा ग्लाइकोलिपिड संश्लेषण के प्रमुख विस्थल है।

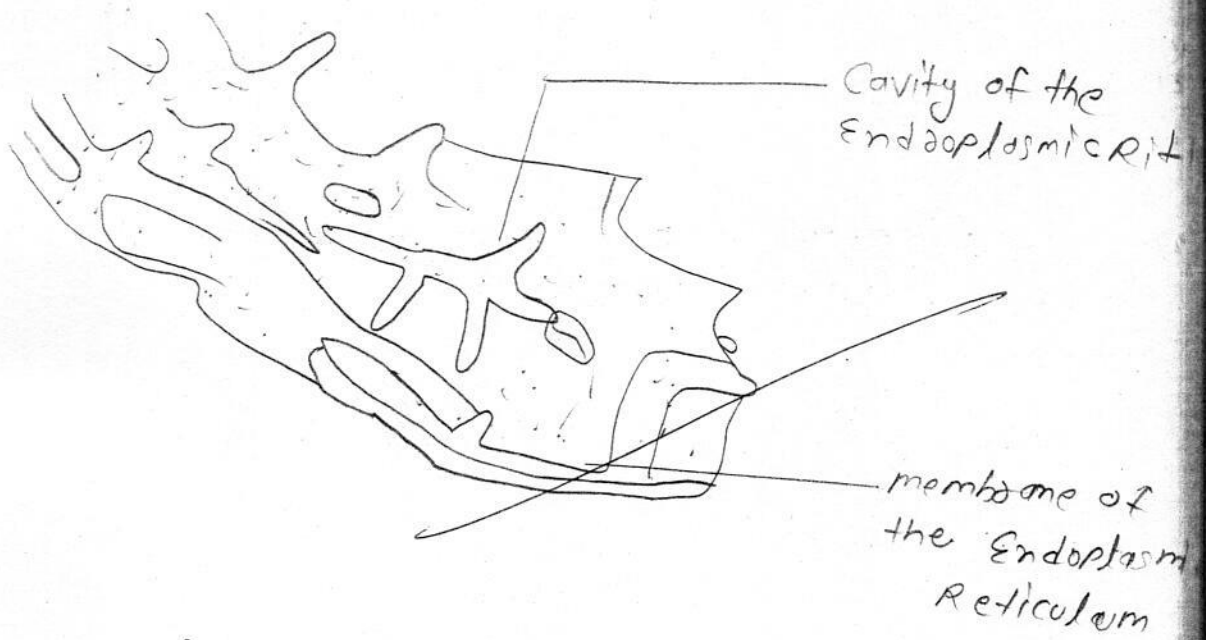
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चित्र:- अन्तः पृष्ठी जालिका

Abhinav
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10. अन्तः प्रद्वी जालिका (Endoplasmic Reticulum)

1. यह चपटे एक सिल्ली आवरित सिस्टर्नी नलिकाओं व आशययुक्त केन्द्रक आवरण की बाहरी सतह से लेकर प्लाज्मा कला तक बिखरी रहती है।
2. इनकी सतह पर राइबोसोम उपस्थित रहते हैं वह रक्षा कहलाते हैं।
3. राइबोसोम द्वारा निर्मित प्रोटीन की सिस्टर्नी के द्वारा स्थानान्तरित करते हैं।
4. कुछ चिकने अन्तर्द्वी जालिका राइबोसोम रहित होते हैं जो लिपिड व स्टीरॉयड के संश्लेषण विस्थल हैं।

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Genetics

एक संकर क्रास : एक जोड़ी विपर्यासी लक्षणों की वंशागति को सुनिश्चित करने के लिए किए गए संकरण को एक संकर क्रास कहते हैं।

● एक संकर क्रास को निम्न प्रकार से दर्शाया जाता है -

लम्बा \times बौना
(TT) (tt)
Ⓣ Ⓣ

(Tt)

♀	T	t
T	TT	Tt
t	Tt	tt

लक्षण पुरुषी अनुपात = लम्बे : बौने
3 : 1

जीन पुरुषी अनुपात = शुद्ध लम्बे : संकर लम्बे : शुद्ध बौने
TT : Tt : tt
1 : 2 : 1

~~Student~~
06/04/24

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द्विसंकर क्रॉस :- जब दो लक्षणों में विपरीत लक्षणों की वंशागति के अध्ययन के लिए संकरण किया जाता है, उसे द्विसंकर क्रॉस कहते हैं।

गोल पीले बीज
(RRYY)

हरे झुरीदार बीज
(rryy)

x
↓
गोल पीले
(RrYy)

स्वनिषेचन ↓

♂\♀	RY	Ry	rY	ry
RY	RRYY	RRYy	RrYY	RrYy
Ry	RRYy	RRyy	RrYy	Rryy
rY	RrYY	RrYy	rrYY	rrYy
ry	RrYy	Rryy	rrYy	rryy

— F₂ पीढ़ी

लक्षण पुरुषी अनुपात -

गोल व पीले : गोल व हरे : झुरीदार व पीले : झुरीदार व हरे
9 : 3 : 3 : 1

~~Signature~~
12/04/24

[Signature]
PRINCIPAL
VIVEK PG COLLEGE
KALWAR, JAIPUR-303706

द्विसंकर क्रॉस :- जब दो लक्षणों में विपरीत लक्षणों की वंशावृत्ति के अध्ययन के लिए संकरण किया जाता है, उसे द्विसंकर क्रॉस कहते हैं।

गोल पीले बीज (RRYY) × हरे झुरीदार बीज (rryy)

↓
गोल पीले (RrYy)

स्वनिषेचन ↓

	RY	Ry	ry	RY
RY	RRYY	RRYy	RrYY	RrYy
Ry	RRYy	RRyy	RrYy	Rryy
ry	RrYY	RrYy	rrYY	rrYy
yy	RrYy	Rryy	rrYy	rryy

— F₂ पीढ़ी

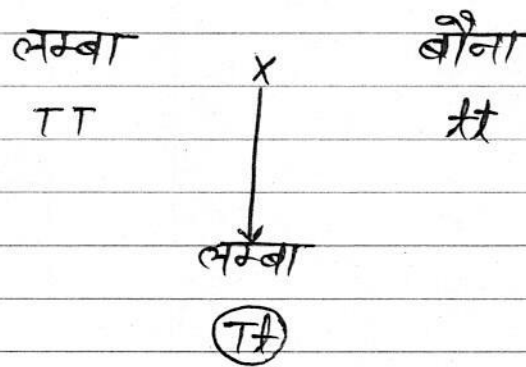
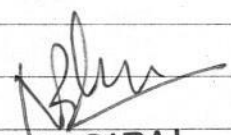
लक्षण पुरुषी अनुपात -
 गोल व पीले : गोल व हरे : झुरीदार व पीले : झुरीदार व हरे
 9 : 3 : 3 : 1

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12/04/24

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 Teacher's Signature

मैण्डल के वंशागति के नियम :-

प्रभाविता का नियम :- इस नियम के अनुसार जब एक जोड़ी विपर्यासी लक्षणों वाले समग्रमजी या शुद्ध पौधों में क्रॉस कराया जाता है तो F_1 संतति पीढ़ी में जो लक्षण अभिव्यक्त होता है उसे प्रभावी लक्षण तथा जो अभिव्यक्त नहीं होता, उसे अप्रभावी लक्षण कहते हैं। इस प्रकार F_1 पीढ़ी में केवल एक ही लक्षण अभिव्यक्त होता है, इसे प्रभावी लक्षण कहते हैं।

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2. पृथक्करण का नियम:- इसके अनुसार F_1 पीढ़ी जो विषम-युग्मजी तथा समलक्षणी प्रभावी होती है के पौधों के मध्य स्वपरागण द्वारा F_2 पीढ़ी प्राप्त की जाती है तो F_2 पीढ़ी में प्रभावी तथा अप्रभावी दोनों लक्षण एकट हो जाते हैं। इनमें प्रभावी लक्षण 75% तथा अप्रभावी लक्षण 25% पौधों में दिखाई देते हैं। इस प्रकार संकर जीव में दोनों विपरीत कारक या जीन साथ-2 रहते हुए संकूपित नहीं होते अथवा अपनी शुद्धता बनाए रखते हैं तथा युग्मक निर्माण के समय एक-दूसरे से पृथक होकर अलग-2 युग्मकों में पहुँच जाते हैं इसलिए इसे पृथक्करण अथवा युग्मकों की शुद्धता का नियम कहते हैं।

(लम्बा) TT × (बौना) tt

(लम्बा) Tt

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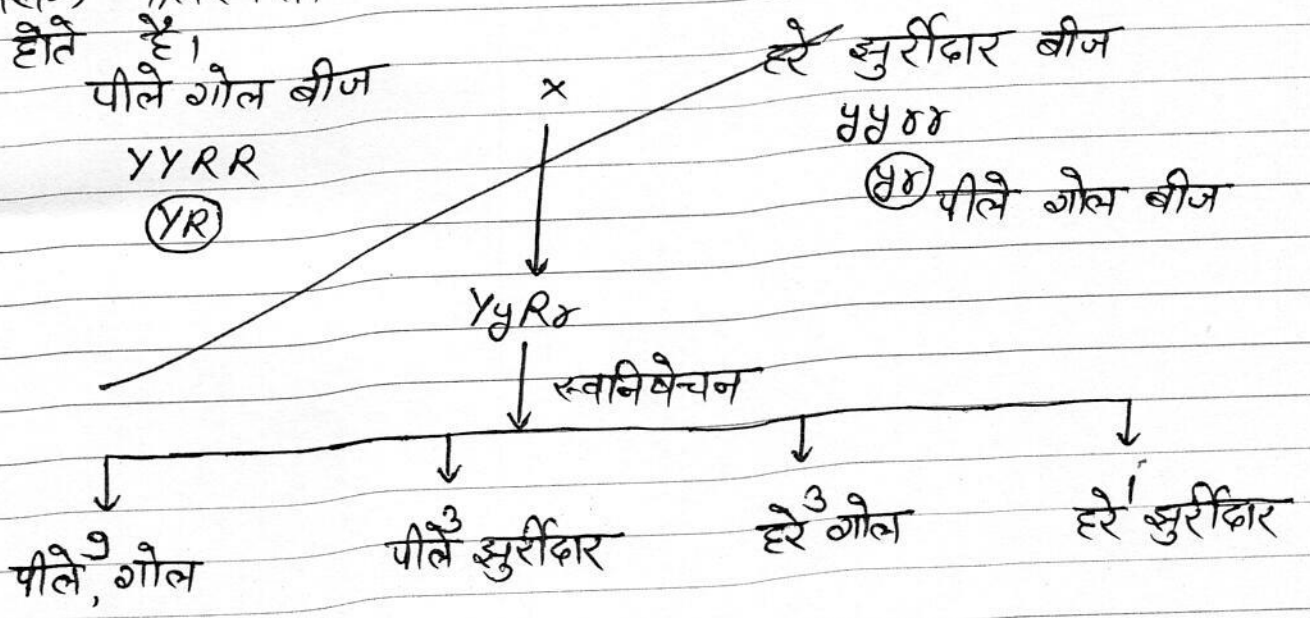
TT	:	Tt	:	tt
1	:	2	:	1

शुद्ध लम्बा	संकर लम्बा	शुद्ध बौना
└──────────┘		
लम्बा 3		: 1 बौना

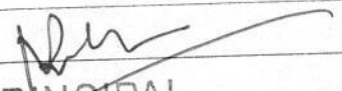
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3. स्वतन्त्र अपव्यूहन का नियम :- इस नियम के अनुसार जब दो या दो से अधिक जोड़े विपर्यासी लक्षणों की वंशागति के अध्ययन के लिए क्रॉस करवाया जाता है तो जोड़े में उप. लक्षणों की वंशागति पूर्णतया एक - दूसरे से स्वतंत्र होती है अर्थात् भ्रूमविकल्पियों के प्रत्येक जोड़े के ऐलील पृथक् होकर विभिन्न लक्षणों के भ्रूमविकल्पियों के साथ स्वतंत्र अपव्यूहन करते हैं। दूसरे शब्दों में यह कह सकते हैं कि अक्रम-2 लक्षणों के कारकों का व्यवहार एक - दूसरे से पूर्णतया स्वतंत्र होता है जिसके फलस्वरूप नए संयोजन या पुनः संयोजन प्रकट होते हैं।



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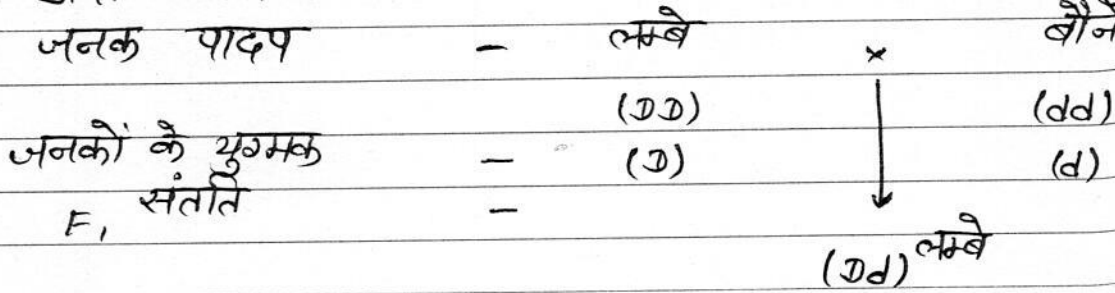

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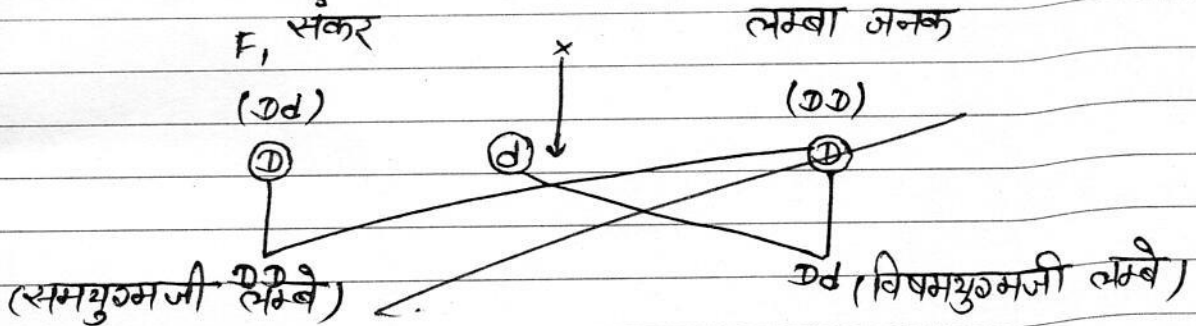
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मैन्डेलिज्म पर अभ्यास

1. मटर के लम्बे (DD) तथा बौने (dd) पौधों के संकरण से प्राप्त पीढ़ी संतति के एक (dd) पौधे का पितृ पौधों से F₁ संकरपूर्वज संकरण किया गया, संकरपूर्वज संकरण से प्राप्त संततियों के जीन पुरुप व लक्षण पुरुप बताइए।



(i) F₁ संतति का लम्बे जनक के साथ संकरपूर्वज संकरण :-



उपरोक्त क्रॉस में प्राप्त सभी पादप लम्बे होंगे अर्थात्

लक्षण पुरुप = 100% लम्बे
 जीन पुरुप = 50% समयुग्मकी : 50% विषमयुग्मकी
 = DD : Dd
 = 1 : 1

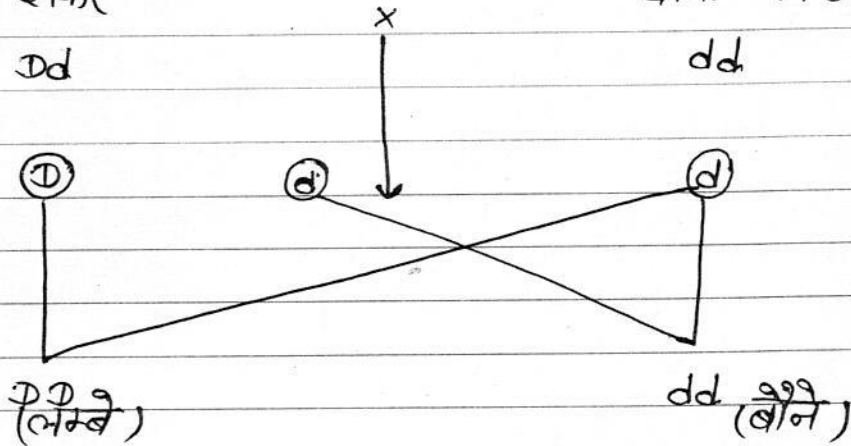
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ii) F_1 का बौने जनक के साथ संकरण :-

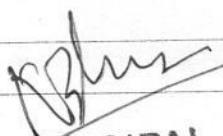
F_1 संकर F_1 बौना जनक

Dd dd



लक्षण प्ररूपी अनुपात - 50% लम्बे व 50% बौने
अर्थात् - 1 : 1

जीन प्ररूपी अनुपात - $DD : dd$
1 : 1


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दिये गये चार्ट पर टिप्पणी लिखें -

1. यह चार्ट मेन्डल के पृथक्करण नियम को दर्शाता है।

2. जब लाल पुष्प के मटर पादप का संकरण सफ़ेद पुष्प के पादप से कराया जाता है तो F_1 संतति केवल लाल पुष्प वाली प्राप्त होती है। क्योंकि लाल रंग सफ़ेद रंग पर प्रभावी है।

3. जब F_1 पीढ़ी के पौधों में स्वनिषेचन कराया जाता है तो F_2 पीढ़ी में लाल पुष्प व सफ़ेद पुष्प दोनों प्रकार के पौधों 3:1 में प्राप्त होते हैं। इसे पृथक्करण का नियम कहते हैं।

4. F_2 संतति में 25% शुद्ध लाल 50% संकर लाल तथा 25% शुद्ध सफ़ेद पुष्पों वाले पौधे होते हैं, जो युग्मकों की शुद्धता को दर्शाता है।

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दिये गये चित्र पर टिप्पणी लिखिये।

1. यह चार्ट टेस्ट क्रॉस को दर्शाते हैं।

2. समयुग्मजी लाल पुष्प वाले पादप का संकरण समयुग्मजी सफेद पुष्प वाले पादप से कराया जाता है तो F_1 पीढ़ी में संकर सुंदर लाल पुष्प वाले विषमयुग्मजी पादप प्राप्त होते हैं।

3. जब विषमयुग्मजी संकर लाल पुष्प वाले पादप का अपभावी शुद्ध सफेद पुष्प वाले जनक पादप से कराया जाता है, तो 50% पौधे संकर लाल पुष्प वाले तथा 50% पौधे सफेद पुष्प वाले प्राप्त होते हैं।

4. टेस्ट क्रॉस हमेशा विषमयुग्मजी विषमयुग्मजी F_1 पीढ़ी एवं शुद्ध समयुग्मजी जनक के मध्य कराया जाता है तो संतति 1:1 में प्राप्त होती है।

~~Signature~~
17/04/24

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Certificate

Name: Heena Agarwal

Class: M.Sc (Final) Zoology

Roll No.: 03

Exam No.: 1859840

Institution: Vivek P.G. College

This is certified to be the bonafide work of the student in the
Zoology Laboratory during the academic

Year 20 23 / 20 24

No. of practicals certified 14 out of 16 in the

Subject of Zoology [Environment & Ethology]

Risham Sharma
.....

Examiner's Signature

[Signature]
.....

Teacher in-charge

[Signature]
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Vivek PG Mahavidyalaya
Kalwar Jaipur-303706

Date: 12/05/24.....

[Signature] Institution Rubber stamp

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Experiment - 1

Aim:- To determine the pH of given water sample

Requirements - Paper strip, universal indicator, water sample, beaker, dropper.

Principle - Generally in nature water is alkaline due to presence of carbonates. water pH keeps on changing due to waste substances released in it by industries mixing of sewage. Production of CO_2 due to photosynthesis change in temp. etc

According to Indian Council of Medical Research (ICMR) pH of potable water should be in between 7 to 8.5 water with pH less than 6.5 or more than 9.2 cannot be used for drinking. If pH is below 4 then water tastes sour and if it is more than 8.5 then it is alkaline in taste water with more pH tends to release metals like zinc, lead, Cd, Cu from pipe lines and may be harmful for domestic as well as industrial use.

pH is a logarithmic scale used to specify the acidity or basicity of an aqueous solution. It is the pH of the base 10 logarithm of the activity of the hydrogen ion. It is measured by the help of 2 electrodes. A galvanic cell is set up to measure the electromotive force between a reference electrode and an electrode sensitive to the hydrogen ion activity when both are immersed in the same aqueous solution.

Procedure - (i) pH paper strip method - It is easiest method

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S.No	Colour of Indicator	Water ph	Water acidity
1.	yellow	4.0	Very intense acidic
2.	greenish yellow	4.5	intense acidic
3.	yellow green	5.0	Acidic
4.	light green	5.5	Medium acidic
5.	greenish blue	6.5	Very weak acidic
6.	Dark blue	7.0	Neutral
7.	Purple	7.5	Alkaline

PH strip


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to determine p^h simply by putting paper strip into a filtered solution of water and one can notice change in the colour of the strip. This colour is matched with the help of colour chart provide on the box of paper strip. Corresponding p^h with colour indicates the p^h of water.

(ii) Indicator solution method :-

Mix equal amount of

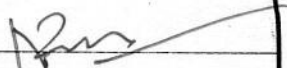
universal indicator in it.

Colour will be seen which is to be matched with the colour chart given on table of bottle note down the water p^h of matched colour

Result :-

A. The p^h of given water sample is 7

B. The result of water sample are 6.5


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Acidity

Sample (50ml)

↓
Methyl orange indicator

↓
yellow

↓
Pink

↓
Methyl orange (-mt)

↓
Titrate with 0.05N NaOH

↓
faint orange

↓
Note reading (A)

↓
continue

↓
add phenolphthalein indicator

↓
Pink


↓
Faint orange

↓
Phenolphthalein indicator (-mt)

↓
titrate with 0.05N NaOH

↓
Pink colour

↓
Note reading (B)


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Experiment-2

Aim :-

To determine the acidity of given water sample

Requirements :-

- (i) Sodium hydroxide (NaOH) - 0.05 N. NaOH molecular weight of NaOH is 40.50 dissolve 2 gm NaOH and dilute to 1000 ml using distilled water.
- (ii) Methyl orange indicator - Dissolve 0.5 gm and dilute to 100 ml distilled water
- (iii) Phenolphthalein indicator - Dissolve 0.5 gm hph in 50 ml water 95% ethanol add 50 ml distilled water add drop wise 0.05 N NaOH in the solution till faint pink colour accordingly.
- (iv) Glassware and other requirements - Beaker, pipette, long dropper, indicator bottles, stand

Principle - To determine acidity by titration, water sample is titrated by NaOH to determine end point. Chemical indicators are used we know that chemical indicators indicate end point by changing colour of solution at particular pH when pH is below neutral pH 7. It is acidic. If sample pH is less than 3.7 then it's titrated by NaOH by adding methyl orange indicator in water sample. The colour indicated end point at 3.7 pH this is known as methyl orange acidity or mineral acidity. Phenolphthalein indicator change colour at 8.3 pH in this case when water sample in terms of hph is



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titrated by NaOH then end point is indicated by
 colour this is called total acidity of solⁿ.
 acidity is measured in between 8.3 pH and 8.3 pH
 is called phenolphthalein acidity or CO_2
 acidity.

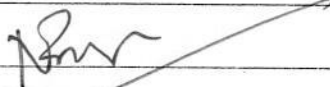
Total acidity - mineral + CO_2 acidity

Procedure :-

Collect sample in plastic or borosilicate
 bottle if should be used for titration within 24hrs
 better to use it as soon as possible take some
 ml of sample water in clean and dry conical flask
 fill the burette with 0.05 N NaOH and note reading
 of its level.

add 2-3 drop of methyl orange indicator in conical
 flask having sample if solⁿ become yellow then it
 mean that methyl orange acidity is 0 (mean pH is
 more than 8.3) if solⁿ turns pink by addition of
 methyl orange then titrate with 0.05 N NaOH drop
 wise burette when pink colour of sample solⁿ
 turn into yellow colour note the reading on burette
 at this end point.

Now add drop of bPH indicator in same solⁿ in
 conical flask and titrate again when solⁿ turns
 into pink then again note reading in burette as
 it will be used in calculation -


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Calculation

$$\text{Acidity of sample I}^{\text{st}} = \frac{(A+B) \text{ N of NaOH} \times 50 \times 1000}{\text{ml of sample}}$$

$$= \frac{(9+0) \times 0.05 \times 50 \times 1000}{50}$$

$$= 450 \text{ mg/l}$$

$$\text{Acidity of sample II}^{\text{nd}} = \frac{(A+B) \times \text{N of NaOH} \times 50 \times 1000}{\text{ml of sample}}$$

$$= \frac{(2.2+0) \times 0.05 \times 50 \times 1000}{50}$$

$$= 110 \text{ mg/l}$$




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Observation table -

S.No.	Water sample taken (ml)	Burette reading of used NaOH sol ⁿ (ml) with methyl orange in titrate	Burette reading of used NaOH sol ⁿ after titration of hPH indicator (ml)	total quantity of NaOH sol ⁿ used in (ml)
1	50 ml	9	0	9
2	50 ml	2.2	0	2.2

Calculation -

(i) Methyl orange acidity -

$$\text{Acidity (mg/l CaCO}_3) = \frac{A \times N \text{ of NaOH} \times 50 \times 1000}{\text{ml of sample}}$$

(ii) Phenolphthalin acidity -

$$\text{(mg/l CaCO}_3 \text{ equivalent)} = \frac{B \times N \text{ of NaOH} \times 50 \times 1000}{\text{ml of sample}}$$

(iii) total acidity -

$$\text{(mg/l as CaCO}_3) = \frac{(A+B) \times N \text{ of NaOH} \times 50 \times 1000}{\text{ml of sample}}$$

Result :- The acidity of given water sample is -

(A) - 450 mg/l

(B) - 110 mg/l

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Alkalinity

Sample (50ml)



Add phenolphthalin



Colourless



Phenolphthalin
alkalinity (mt)



Pink



titrate with 0.1 N
HCl



Colourless



Note reading (A)



Continue



add methyl orange



Pink



methyl orange
alkalinity (mt)



yellow



titrate 0.1 N HCl



Pink orange



Note reading (B)



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Experiment = 3Aim :-

To determine the alkalinity of given water sample

Requirements :-

(i) 0.1 N HCl acid - dilute 12 times 12 N of concentrated HCl means mix 8.4 ml of HCl in distilled water and make it 100 ml or make 800 ml solution by adding distilled water into 12 N HCl this is 0.1 N HCl take 100 ml of this 0.1 N HCl and add distilled water to make it upto 1000 ml this is 0.1 N HCl.

(ii) HPH indicator - dissolved 0.5 gm HPH into 50 ml of 95% ethanol or ethyl alcohol add to few drops of CO₂ free 0.05 N NaOH until the solⁿ become light pink

(iii) Methyl orange indicator - Dissolve 0.5 gm methyl orange in 100 ml of distilled water to make the indicator

Principle :-

Alkalinity in water sample is due to carbonate, bicarbonate and hydroxyl ion their quantity can be measured by titrating it with strong acid if solⁿ is sufficient. carbonate is 4mt then such solⁿ turns pink to pH which is more than 8.3 if this solⁿ is titrated with acid then at pH 8.3 all carbonate turn into bicarbonate and pink colour disappear and solⁿ become colourless.

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Due to alkalinity carbonate of all the 3 forms of buffer system can be measured separately but for convergence alkalinity is show as equivalent to mg/l or ppm calcium carbonate therefore if total alkalinity of solution is 125 mg/l then it mean that if in pure water 125 mg/l CaCO_3 is mixed and as result the amount of alkalinity produced due to this is the same alkalinity of given solution

Procedure :-

Collect sample to titrate the sample once 100 ml of sample is required so accordingly collect sample in plastic or borosilicate bottle

→ take 50 ml of sample in clean & dry conical flask and mix 2-3 drop of phenolphthalein indicator if solⁿ remain colourless then it mean that hph alkalinity in the sample is 0.

→ If sample turns pink by adding phenolphthalein indicator then titrate it with 0.1 N HCl filled in burette if solⁿ become colourless then note to the end point

→ if it is colourless by adding hph then add alkalinity is 0 2-3 drop of methyl orange indicator and bicarbonate alkalinity is 0 and in this condition hph alkalinity is the total alkalinity are equal if colour of solⁿ become yellow then titrate it with 0.1 N HCl until the colour turns into light pink.

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Calculation :-

$$\underline{\text{Alkalinity of sample I}} = \frac{(A+B) \times N \text{ of HCl} \times 1000 \times 50}{\text{ml of sample}}$$

$$= \frac{(0 + 1.5) \times 0.1 \times 1000 \times 50}{50}$$

$$= 150 \text{ mg/l}$$

$$\underline{\text{Alkalinity of sample II}} = \frac{(A+B) \times N \text{ of HCl} \times 1000 \times 50}{\text{ml of sample}}$$

$$= \frac{(0 + 7.6) \times 0.1 \times 1000 \times 50}{50}$$

$$= 760 \text{ mg/l}$$




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Observation table -

S.No.	S. quantity in ml	with burette reading in (ml)					
		HPH indicator			Methy orange		
		initial	Final	total (A)	Initial	final	total (B)
1.	50 ml	0	1.5		0	1.5	1.5
2.	50 ml	0	7.6		0	7.6	7.6

Calculation - Phenolphthalin alkalinity -

$$(PA \text{ as } CaCO_3 \text{ mg/l}) = \frac{A \times N \text{ of HCl} \times 1000 \times 50}{\text{ml of sample}}$$

methyl orange alkalinity

$$(MO \text{ as } CaCO_3 \text{ mg/l}) = \frac{B \times N \text{ of HCl} \times 1000 \times 50}{\text{ml of sample}}$$

$$\text{Total alkalinity} = \frac{A+B \times N \text{ of HCl} \times 1000 \times 50}{\text{ml of sample}}$$

Result - The alkalinity of given water sample is
 a. 150 mg/l
 b. 760 mg/l

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Free CO₂

Sample (50ml)



Add phenolphthalein



Pink

CO₂ amt

colourless

titrate with 0.0
N NaOH

Pink



Note reading



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Experiment - 4

m :-

To determine the free CO_2 of given sample requirements :-

(i) 0.05N NaOH - prepare 2 l solⁿ by dissolve 10 gm of NaOH in distilled water strain this solⁿ through standard glass filter so that Na_2CO_3 is filtered remain in filter 1N NaOH obtained through this can be stored in airtight bottle

(ii) Phenolphthalein indicator - Dissolve 500mg pH in 50 ml 95% ethanol add 50 ml of distilled water then add 0.05N CO_2 free NaOH solⁿ dropwise until the solution turns light pink -

Principle :-

Na_2CO_3 is formed by the neutralization of carbonic acid. After the neutralization solⁿ become alkaline this can be measured by using pH indicator which produce pink colour at 8.3 pH method to estimate free CO_2 - titrate by NaOH titrate by Na_2CO_3

Procedure - Collect water sample carefully in 250-300ml bottle or 400 ml nessler tube while collecting water bubbles should not be there in water should be disturbed more if water sample is to be collected from depth then kammerer water sampling bottle is appropriate to use.

Calculation :-

$$\text{free CO}_2 \text{ of Sample I}^{\text{st}} = \frac{A \times N \text{ of NaOH} \times 1000 \times 44}{\text{ml of Sample}}$$

$$= \frac{12.2 \times 0.05 \times 1000 \times 44}{50}$$

$$= 536.8 \text{ mg/l}$$

$$\text{Free CO}_2 \text{ of Sample II}^{\text{nd}} = \frac{A \times N \text{ of NaOH} \times 1000 \times 44}{\text{ml of Sample}}$$

$$= \frac{8.1 \times 0.05 \times 1000 \times 44}{50}$$

$$= 356.4 \text{ mg/l}$$

✓

- take 100 ml of water sample in clean & dry conical flask add few drops of HPH indicator in it solⁿ are pink
- if sample remain colourless then it should be titrate by 0.05 N NaOH filled in burette.
- Note the end point when the solⁿ turns pink in colour. Turn off the burette knob and write down the reading of volume of 0.05N NaOH.

Observation table -

S.No.	Volume of Sample (ml)	Burette reading (ml)			mean
		initial	final	total	
1.	50 ml	0	12.2	12.2	
2.	50 ml	0	8.1	8.1	

Calculation -

$$\text{Free CO}_2 (\text{mg/l}) = \frac{A \times N \text{ of NaOH} \times 1000 \times 44}{\text{ml of sample}}$$

Result - The free CO₂ of given water sample are -

A. 536.8 mg/l

B. 356.4 mg/l

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Salinity (Chloride)

Sample



Potassium dichloride
indicator



Yellow colour



titrate with
0.02 N $AgNO_3$



Milky colour




Red brick colour



Note reading




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Experiment - 5

Aim :-

To determine the salinity of given water

Sample.

Requirements :-

(i) 0.02N silver Nitrate - Dissolve 3.49 AgNO_3 in water and make it upto 1000 ml by adding distilled water. Keep this solⁿ in coloured glass bottle or normal transparent bottle wrapped with carbon and opaque paper for AgNO_3 in solⁿ react due to sunlight, and become inactive

(ii) 5% potassium ^{Chromate} Chloride (K_2CrO_4) - dissolve 5 gm K_2CrO_4 in 100 ml of distilled water

Principle - Chloride is too soluble in water so it is very difficult to remove it is very difficult to remove it from water. Chloride gives salty taste to water at 250 to 500 mg/l but upto 1500 mg/l they are not harmful. According to Indian Council of Medical Research (ICMR 1975) the amount of chloride in potable water should not be more than 1000 mg/l. Excess of chloride in water are harmful for building too because these react with calcium amt in concrete to form calcide and lower the strength of concrete so water having mg/l₂ should not be concrete used in boilers. Chloride amt in water react with silver Nitrate and forms white precipitate of silver chloride.

92
Calculation :-

$$\text{Salinity of sample I}^{\text{st}} = \frac{A \times N \text{ of AgNO}_3 \times 1000 \times 35.5}{\text{ml of sample}}$$


$$= \frac{7.1 \times 0.02 \times 1000 \times 35.5}{50}$$

$$= 100.82 \text{ mg/l}$$

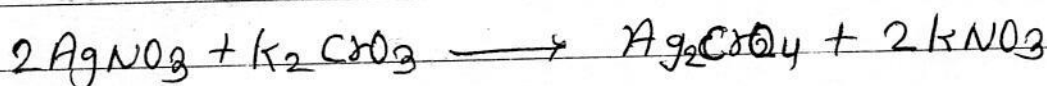
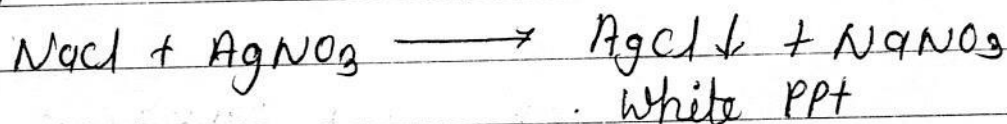
$$\text{Salinity of sample II}^{\text{nd}} = \frac{A \times N \text{ of AgNO}_3 \times 1000 \times 35.5}{\text{ml of sample}}$$

$$= \frac{8.7 \times 0.02 \times 1000 \times 35.5}{50}$$

$$= 123.55 \text{ mg/l}$$


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When all chloride are precipitated then silver nitrate is found in free state in the solⁿ it can be confirmed by using potassium chromate indicator because this indicator forms brick coloured silver chromate as end point when titrated -



Procedure - Collect sample in plastic or borosilicate bottle for one time titration 50ml sample is required. Sample can be kept for 7 days for measuring chloride.

→ take 50ml of sample in a conical flask add 2ml of potassium chromate indicator into it colour change from yellow to milky if flask is not shaken continuously then you see the red colour but on shaking it disappears but at some point red colour comes and if does not disappear on shaking this is the end point.

Observation table -

S.No.	Volume of Sample	Burette reading (ml)			Mean
		initial	final	total	
1.	50ml	0	7.1	7.1	
2.	50ml	0	8.7	8.7	

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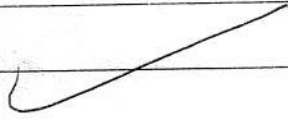
Calculation -

$$\text{Chloride (mg/l)} = \frac{A \times N \text{ of } AgNO_3 \times 1000 \times 35.5}{\text{ml of sample}}$$

Result - The salinity of the given water sample

are - A - 100.82 mg/l

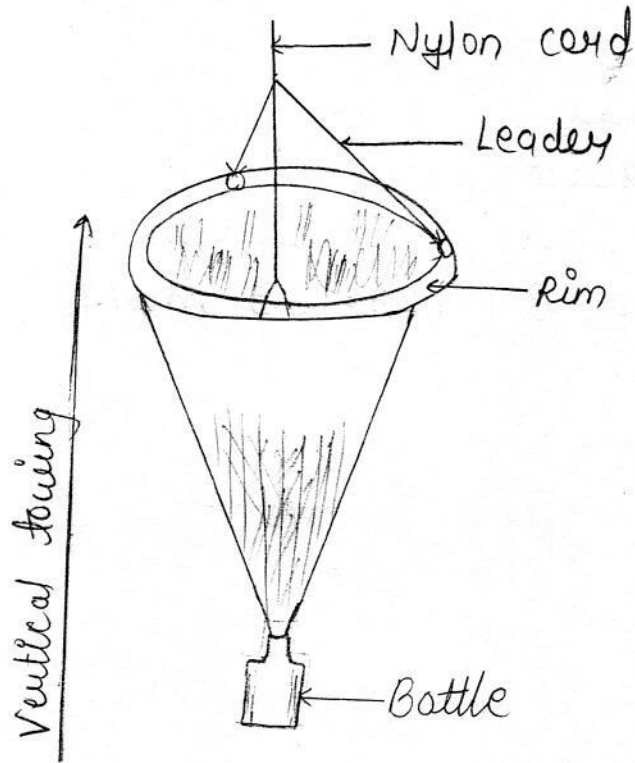
B - 123.55 mg/l



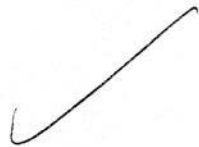

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Plankton net




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Experiment - 6

Aim :-

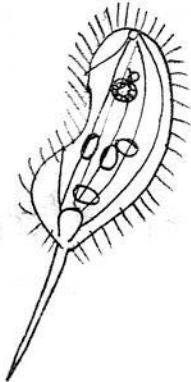
Collection, isolation and identification of plankton
(phyto and zoo plankton)

Plankton Collection - Plankton can be studied properly if they are collected and method for collection are as follows -

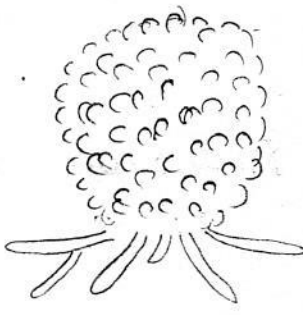
(i) Bottle sampling - Sample is collected by immersing bottle in water so that plankton come into the bottle along with water this method is easy but has disadvantages too firstly plankton try to escape & Secondly their no is too less in the sample taken so one has to collect sample 3 to 4 times to study them properly beaker can also be used instead of bottle.

(ii) Plankton Net - Plankton net is used to collect plankton it is made up of thin silk or nylon fibre its mesh size is fixed. Net is made up of a circular iron ring attached to handle the ring is tied with belting silk cloth having different mesh size out of which 60µm most common net is used with slow motion (10cm/sec) depth of place where sample water for plankton is collected can be measured by using plankton net collect plankton by cylindrical column of water we know that volume of a cylinder can be calculated as

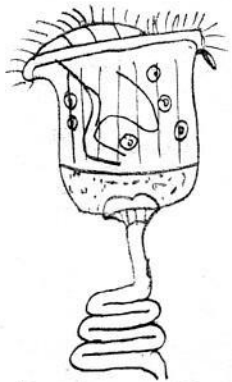
$$V = \pi r^2 L$$



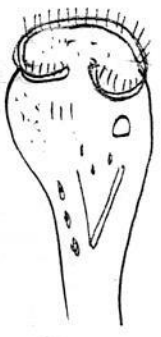
Hyalotheca



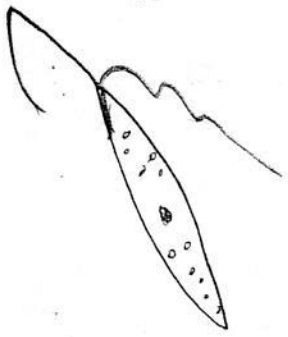
Diffugia



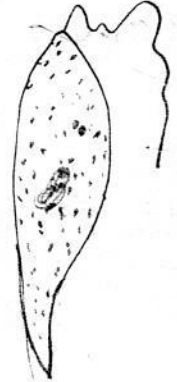
Vorticella



Stentor




Paramecium



Euglena

Organisms of protozoa


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Where $\pi = 3.14$, $r =$ radius of rim of net, l is length covered by net through this method large no of plankton are collected.

(B) Fixation and preservation of plankton - To identify and count the plankton it is essential to keep them for long through try to study them live with the help of microscope for better result. plankton are fixed using following solution -

(i) Lugol's solution - This solⁿ preserve plankton as well as stain them too but they cannot be preserved in this solution for long plankton sample preserved in lugol should be kept in dark.

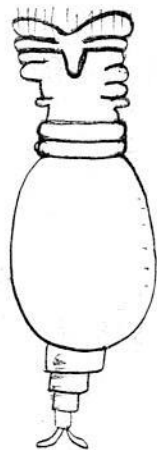
(ii) formalin - this solution is simple & easily available 4% formalin is adapt for preservation formalin is saturated solⁿ for formaldehyde in water.

Qualitative study of plankton - plankton are affected by the physical & chemical factors of water. Some are found in clean water & some can survive in the most polluted water too. Some plankton live in soft water & some live in salty water. Some plankton inform about the pollution level of water in which they live and therefore called bio indicators for eg. Euglena, Oscillatoria.

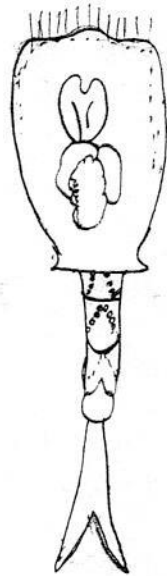
(i) Protozoa - Organism are one called some are colonial; chlorophyll is cont only in Euglena for locomotion cilia flagella or pseudopodia may be cont



Rotaria

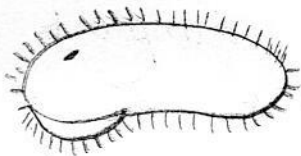


Philodina

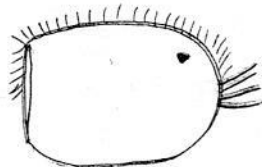


Scarcidium

Organism of Rotifera

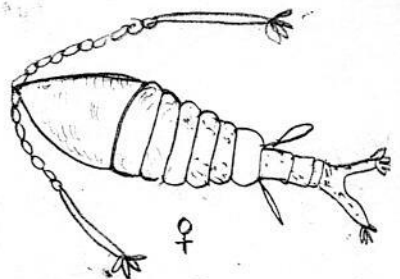


Heterocypis



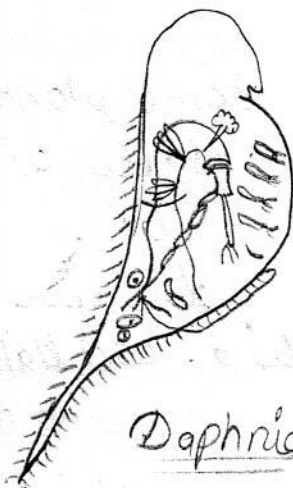
Centrocypis

Organism of Ostracoda

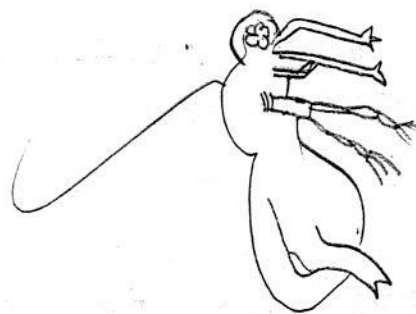


Spicodiantomus

Organism of Copepoda



Daphnia carinata



Moina (male)

Organism of Cladocera

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- (ii) Rotifera - generally worm like organism for locomotion one or two circlet of cilia are present on body called corona.
- (iii) Ostracoda - Organism with jointed appendages external skeleton present. this exoskeleton (shell) is bivalve type like in unio they can be retracted in shell if required.
- (iv) Copepoda - Body divided into 2 parts jointed appendages antenna non-branching
- (v) Cladocera - Body division are not clear. Body is flat and posterior part is covered by carapace locomotion is by second antenna.


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Experiment - 7

Aim:-

To study the food preference of stored insect pest (*Tribolium*) in whole grain

Requirement:-

Tribolium, multicompartment box, types of whole grain, wheat, rice, barley, pearl millet

Principle - It can be an interesting subject to study the food preference of insect pest. Take a container which has places where different kind of food material can be kept and they won't get mixed. This container should be closed so that other insect can't go inside and insects which are inside cannot go out.


A large numatic trough or dry fruit tray can be used for this purpose. Put different material in different chamber and note the no. of insect in each chamber. Repeat the experiment in different chamber and note the no. of insect in each chamber. Repeat the experiment with same food material like *Somaling flour*, all purpose flour, wheat, maize etc. are the product of basic substances. Wheat. Keep these product and then test which product are like more by insect. This can be done with different species also. It can be checked that if disliked food is given, then what happens. These are proved to be informative.

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Calculation:-

Substance	live	dead
Wheat %	$\frac{34}{62} \times 100 = 55.73$	$\frac{0}{62} \times 100 = 0$
Bajra %	$\frac{20}{62} \times 100 = 32.78$	$\frac{1}{62} \times 100 = 1.63$
Rice %	$\frac{5}{62} \times 100 = 8.19$	$\frac{1}{62} \times 100 = 1.63$
Pearlmillet	0	0


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as well as interesting by these studies we can know the precaution to be taken in storage of food and how food substances can be kept safe by using what kind of product this way many experiment can be performed.

Procedure:- first of all multicompartement box in which different types can be kept but they do not mix with each other. Tribolium is left in the middle space of the box around which different types of grain are placed.
 → close the box out of which other insect can't go inside and the insect could not come out.
 → give the no. of tribolium in every type of substances on the next day.

Observation table -

S.No	grain	tribolium		total
		live	dead	
1.	Wheat	34	0	
2.	Rice	5	1	
3.	Bowley	20	1	
4.	Pearl millet	0	0	62

Result - The food preference tribolium in grain wheat

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Experiment - 8

Aim :-

To study the food preference of stored insect pest (*Tribolium* sp.) in powder grain.

Requirements :-

Tribolium, multi compartment box types of whole powder grain, wheat flour, maize flour chickpea flour (besan) suji, maida etc.


Principle :-

It can be an interesting subject to study the food preference of insect pest take a container which have places where different kind of food material can be kept and they won't get mixed this container should be closed so that other insect can't go inside which are inside cannot go out. A large numeric trough or deep fruit tray be used for this purpose put different material in different chamber and note the no. of insect in each chamber repeat the experiment with same food material the something flour all purpose flour wheat maida etc are the products of basic substances wheat keep these product and then test which product or products are like more by insect this can be done with different specie also it can be checked that if disliked food is given then what happen these are proved to be informative as well as interesting by these studies we can know the precaution to be taken in

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Calculation :-

Substance	live	dead
wheat powder & grain	$\frac{4}{70} \times 100 = 5.71$	$\frac{1}{70} \times 100 = 1.42$
Suji	$\frac{14}{70} \times 100 = 20$	$\frac{1}{70} \times 100 = 1.4$
maize powder	$\frac{29}{70} \times 100 = 34.28$	$\frac{1}{70} \times 100 = 1.4$
Besan	$\frac{9}{70} \times 100 = 12.85$	0
maida	$\frac{16}{70} \times 100 = 22.85$	0


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Storage of food and how food substances can be kept safe by using what kind of products this way many experiment can be performed.

Procedure - Multicompartment box add & put different types of grain flour in all compartment and there is a circle in the middle then cover that box overnight and break it then after observing if the next day count the % of tribolium % in the flour the flour which contain more tribolium is a animal food.

Observation table -

S.No	food grain	No of tribolium		total
		live	dead	
1	Wheat powder	4	1	
2	Suji	14	2	
3	maize powder	24	1	
4	basan	9	0	
	maidg	16	0	70

Result - The food preference tribolium in powder maize.

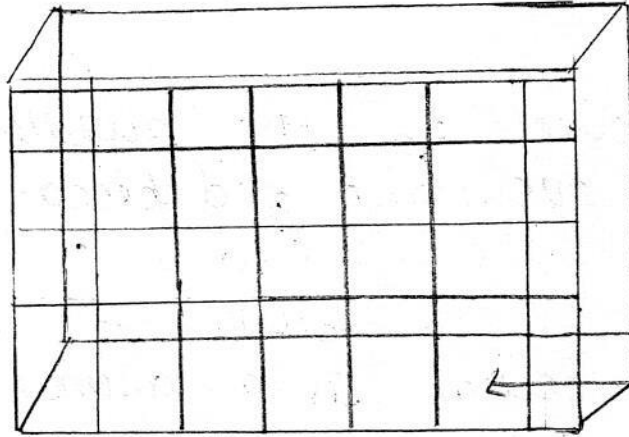
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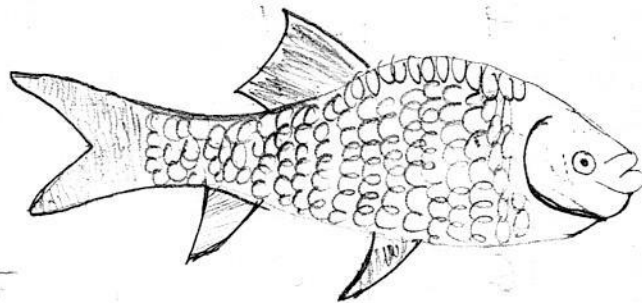
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Squares



Aquarium with square & fish


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Experiment - 9

Aim:-

Observe of Effect of Toxicants on Movement of fish.

Material:-

Chana punctatus (Ophiocephalus), a cat fish or any other medium sized easily available fish, aquarium (3 ft x 1.5 ft), permanent marker, stop watch, a toxicant.

Method:-

Draw vertical and horizontal lines on the outer sides of the aquarium with a thick water proof marker pen, at least three inches apart, on all five sides of the aquarium (four sides and bottom). This way squares will be formed. To see the effect of a toxicant it can either be injected, or given orally by force feeding with the help of tube, or mixed in water (the dose has to be decided in consultation with an expert). Take six fish, keep three as control i.e. untreated and three as experimental i.e. treated.

Start the preliminary experiment, watch their movement, practice and count how many squares a fish enters in two minutes (use the stop watch, this interval can be changed - if the fish is very active then time interval can be reduced but if the fish is slow then it can be increased, and once decided then it has to be kept constant through out the experiment). Once you have practiced, then take the actual observations. First on untreated fish then on treated fish.

Number of days can be decided by the experimenter. Compare the means, if you wish to analyze the data statistically then increase the number of fish and days of observations, then analyze data using 't' test.



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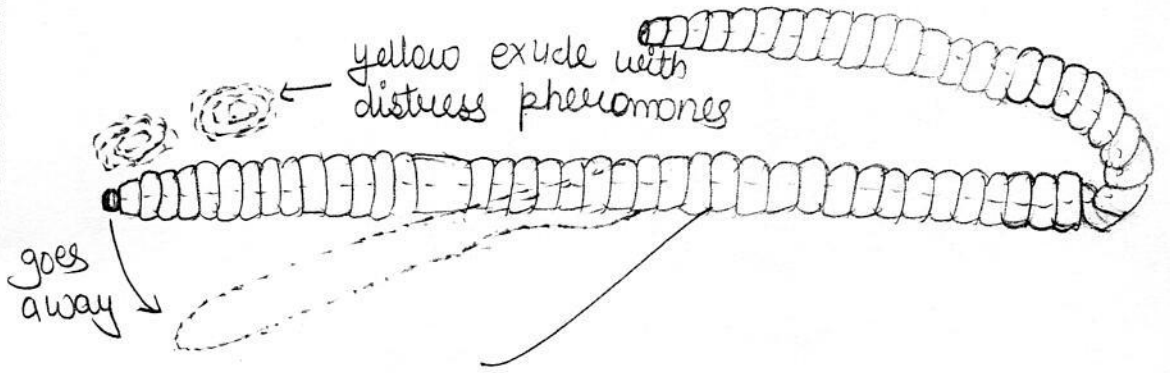
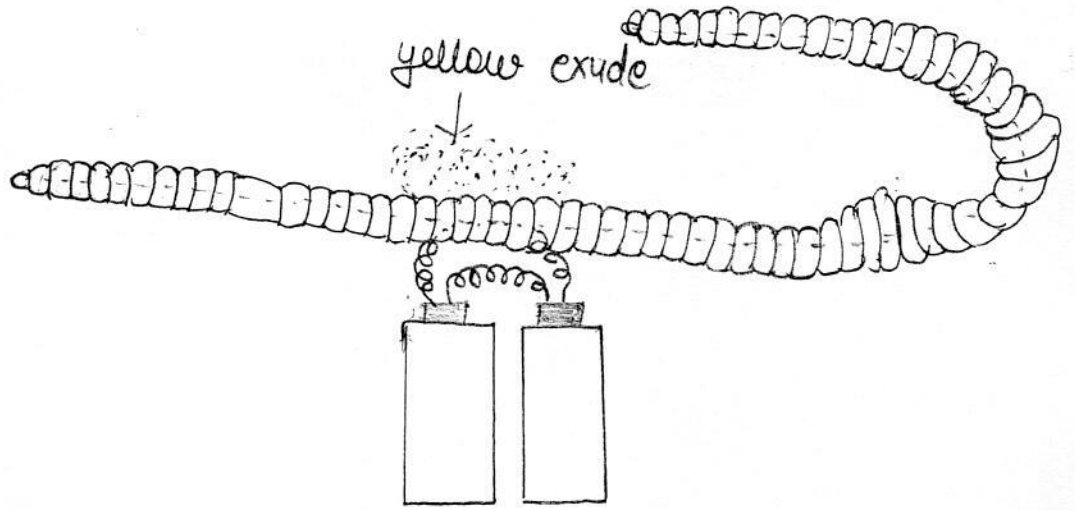
Note how many squares untreated normal fish covers
in compare it with treated fish and evaluate the affect
toxicant on movement of the fish.


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Mild current is given to earthworm
Earthworm goes away from distress
pheromones


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Experiment - 10

n:-

Observe of communication in Earthworm by Pheromone production:-

Visual and auditory stimuli can not carry message earthworm since they do not have visual or auditory organs. Chemical stimuli can be used for communication but this would be used to a time when worms are in proximity. However, earthworms depend most on chemical communication.

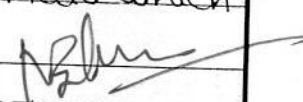
Material:-

● Blunt forceps, paper-towels, scissors, waxed paper, or glazed tiles (which are usually put on top of the tables in labs) size D batteries connected in series with a wire. A strong solution of table salt and water. Live earthworms (earthworms should be handled carefully and gently).

Procedure:-

At first cut strips of paper towels 2 cm wide, soak them in salt solution and arrange them in a square on a piece of waxed paper. Then put two earthworms in the centre of square and then observe their response of each other, to the waxed paper and to salt solution.

● Put another earthworm on the waxed paper or a tile give mild shock to earthworm by touching it briefly with the wires coming from the two size D batteries. The shock will cause the worm to exude yellowish coelomic fluid from the grooves between the segments. Remove this earthworm and put another worm in this fluid and observe its reaction towards the yellow fluid which exuded out of the first worm.


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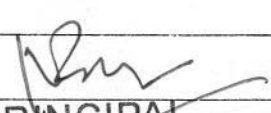
ervation:-

If the response is negative, the worm will jerk head up and move back, away from the yellow exude, thus showing that the coelomic fluid given out by the worm due to toxic shock was a repelling pheromone. If the experiment is repeated 100 times then 95% of times earthworms will show negative response.

As an additional exercise you can study ants also. Follow the trails. Watch ants communicating with each other by tapping antennae.

ult:-

The earthworms communicate by pheromones which are the chemicals expelled from an organism and elicits a response in a conspecific organisms.


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Experiment - 11

aim:-

Observe of learning by Trial and error in Rat with the help of Hebb - William Maze.

Introduction:-

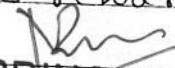
Learning represents change in behaviour, it is different than instinct learning because it is not genetically controlled and is flexible.

Learning is a relatively permanent modification of behaviour due to motivation. It is associated with reward and punishment. However, in the absence of reward or punishment it can be extinct.

Learning has been classified differently by various scientists. Thorpe's (1963) classification is most widely used:

- 1) Flexible. (i) Habituation; (ii) Classical conditioning;
- 2) Trail and Error; (iv) Latent; (v) Discrimination
- 3) Restricted. (i) Imprinting.

When animals are motivated by thirst, hunger, sex or fear they show restlessness, and exploratory of appetitive behaviour during the course of which it performs spontaneously a variety of motor patterns viz - sniffing, walking and looking around. One of these patterns is followed by reinforcement e.g. a hungry animal while exploring the surrounding receives food and if this association is repeated the animal learns to perform a pattern regularly to that particular situation. Animals learn to eliminate behaviour, which lead to no reward and increase the frequency of behaviour that is rewarding by trial and error.

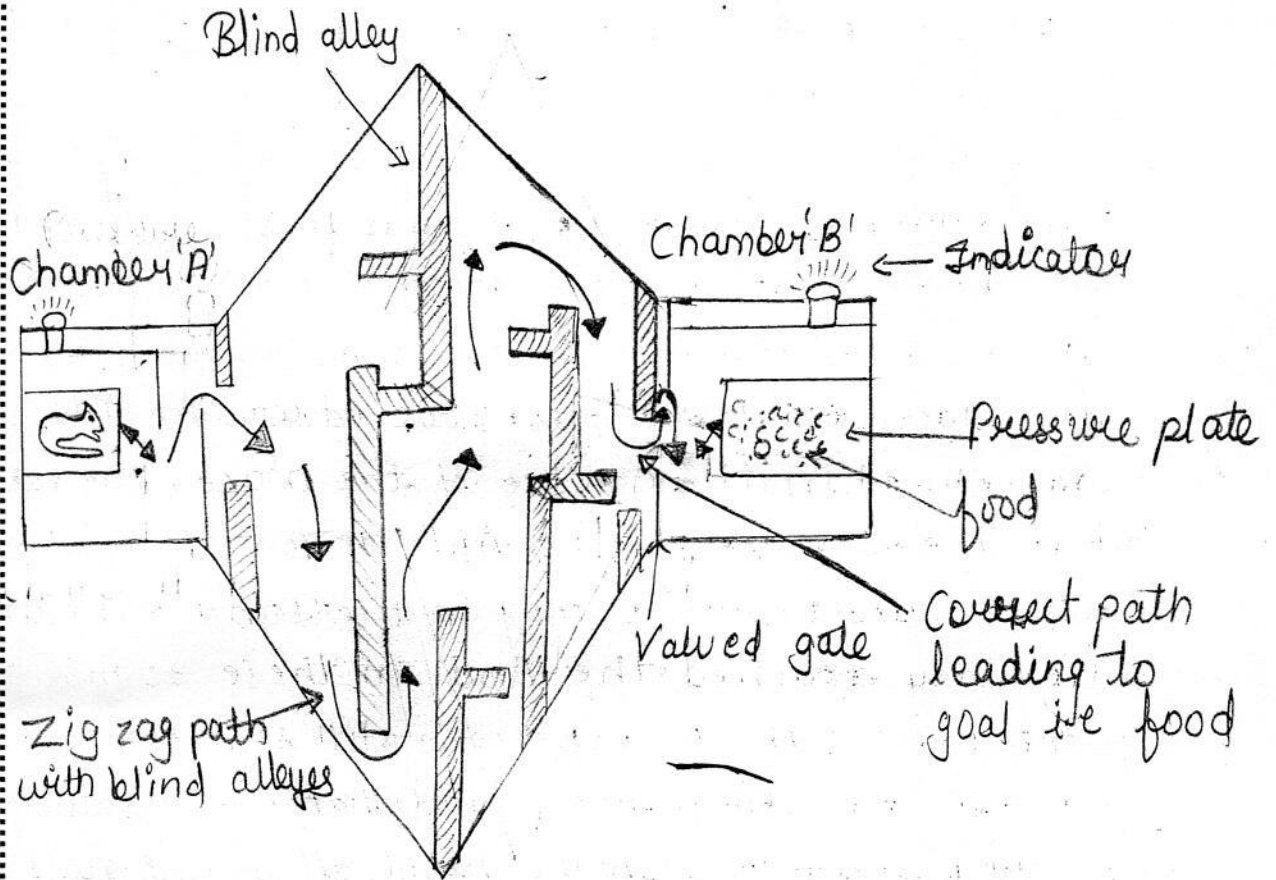

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Network of few lines and paths is called a "simple maze". A system of complex passages where only one leads to the goal while others come to a dead end is called a "complex maze". Maze experiments have played an important role in studying the learning behaviour. An important role in or measure for learning is the number of trials which an animal takes to find the goal without error that is, without entering the blind alley - learning of complex path was first studied by Small (1899-1900). He built a 6' x 8' research maze and used it as an instrument in research. It was constructed by a wire mesh and placed on sand dust covered wooden floor. Some food was placed in the centre of the maze. The rats were placed at the entrance working independently, they explored the whole maze trying to dig in the sand dust or bite the wire. They reached the food in these 30 minutes. On subsequent trials, the time became still shorter and the error were decreased. It was noted that when rats were not hungry, they would play in the blind alley but quickly dash to the goal box when hungry.

Material:-

One cage, 4 white male rats, maze (readymade maze are available with firms selling instruments for psychological tests, a maze can be fabricated using the design given below), stopwatch, water and food for the rats. Before any experiment is carried on animals. It is necessary for the students to get familiarity with the animal. This familiarity can be achieved by observation and handling of the animals.



HEBB WILLIAM MAZE

Hebb William maze with two chamber & zig zag path

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Methods :-

(1) Check all the light points.

Mark 4 male rats (the number can be increased) differently (use a permanent marker, mark one on head, one on trunk, three at tail joint and don't mark the fourth one).

Starve the animals 24 hours prior to reading, but give them water.

Keep a starved rat in chamber A the bulb lights.

Keep nice odorous food in chamber B.

Start the stopwatch when the rat comes out of the chamber A, close the door of chamber A.

Whenever rat enters a blind alley note it as an error. Note the time when the rat reaches chamber B, the bulb lights.

By this time we will have total number of errors and total time taken.

Repeat the same with remaining rats.

After training put the rats back to the cage. Give them food.

Keep the cages clean and repeat the procedure after every second day after starving them. Day one you will starve the rats, second day take reading, third day give, fourth day reading so on and so forth.

Result :-

With subsequent trials, the time taken to reach chamber B, and number of errors is reduced indicating that rats learnt the maze by trial and error.

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Experiment - 12

nº:-

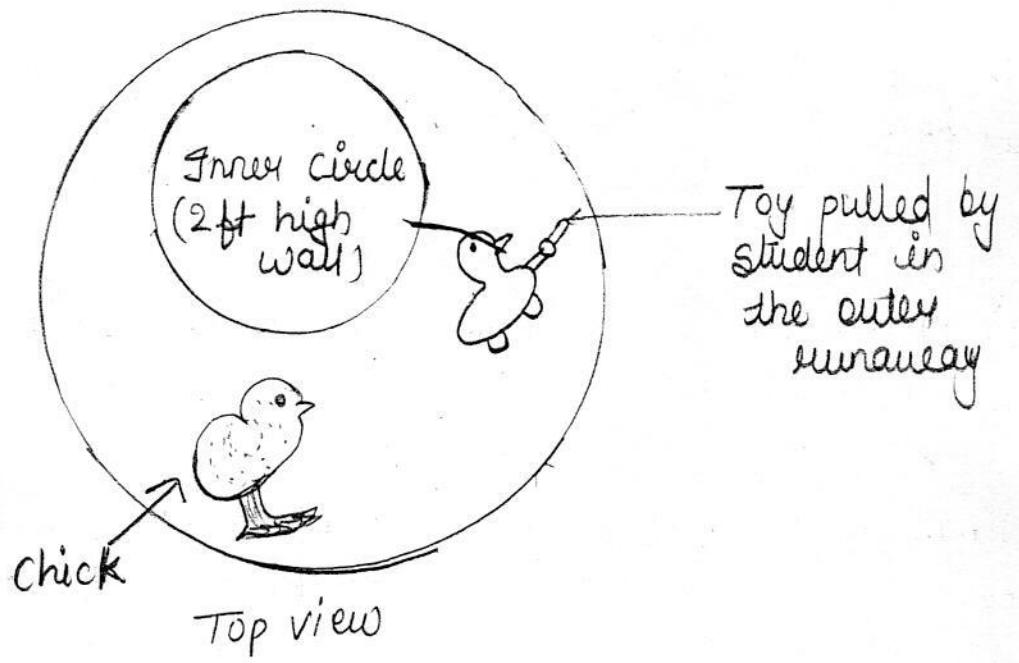
Observe of imprinting in chicks During critical period.

Introduction:-

Imprinting is a learning process that occurs at remarkably early age. Konrad Lorenz (1935) was the first to notice this type of learning and he coined the term "imprinting". He was a naturalist, lived on his farm with a variety of animals. For his work on imprinting he was given a Nobel Prize in 1973. Konrad Lorenz's observation and interpretations on imprinting are remarkable. He mainly worked with graylag geese and Jackdaws. The easiest way to explain the idea of imprinting is to how Konrad Lorenz's first studied it. On his farm, he found an abandoned clutch of graylag goose eggs. He collected them and put them in an incubator, after hatching the goslings started following Lorenz wherever he went. Then he decided to do a simple experiment. This time he deliberately picked half of the eggs from a clutch, he left rest for the mother to incubate. When the chicks hatched, he got all of them together under a box, when they were released half of them followed to mother and half came to Lorenz. Lorenz called this phenomenon *Imprinting*, which is a German word meaning "stamping in" because the gosling were stamped an impression of the particular parent object with which they had their first social experience. Lorenz also noticed that the initial social experience must take place during a critical period early in life. According to Lorenz imprinting is a unique form of learning because it

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Circular runway showing a chick following a toy

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Takes place only during a brief sensitive period early in life. has stability, generally lasting for the rest of the animal's life and

influences the animal's choice of parents and sexual partner. Newly hatched precocial bird (chicks move soon after hatching) ducks, geese, chickens turkey follow a wide range of object move before them, say within 48 hours after hatching, young birds following response will continue for many days or weeks (species specific). However, if following behaviour is not elicited during the early period, it generally not be produced at a later age. The following or strength of imprinting is increased if the stimulus makes noise as it moves. In nature the chicks always see their parents when they hatch that noise and move around, so the chicks start following them. To study the phenomenon of imprinting in newly hatched chicks have to be procured from the poultry farm.

Material-

Newly hatched chicks, two circular aluminium enclosure, fitting inside another a duck toy. The chicks should be kept in separate compartments (approximately $15 \times 10 \times 10$ cm) to maintain visual and tactile isolation (if the chicks can see each other they get imprinted with each other). The birds should be 18 hours old (post hatch) for maximum responsiveness.

Method:-

Take one chick at a time for training. Put one chick in a circular runway. Put the toy duck in front of the chick and pull it slowly at a speed comparable to that a normal

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chick, the toy should also make a noise.

If the model is made to move and stop the proportion of following will increase. Expose each chick individually to the model for 20 minutes. Make your observations from behind a curtain or screen. Record the number of minutes the chicks actually spend following the model. Train the chicks every day and for a week.

If you have time and space train chicks to follow you, they will get imprinted to you.

Result :-

After 7 days, test the chicks for imprinting. Take them out in an open but secure space and drag the toy duck and see how chicks follow this mother model. What actually you would see that some times they are following sometimes not. You leave the room, come back after an hour, you will find all the chicks near the toy. The chicks have got imprinted with this toy.

Ayasha
16.1.24

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Experiment - 13

Object - To determine dissolved oxygen of given water sample.

Apparatus - Titration flask, titration tube, manganous sulphate solution, Alkali iodide azide reagent, concentrated sulphuric acid, sodium thiosulphate solution & starch solution

Principle - If manganous sulphate is added to the sample containing alkaline potassium iodide manganous hydroxide is formed which is oxidised by the dissolved oxygen on addition of strong sulphuric acid is added the basic manganic oxide liberates iodine equivalent to that of dissolved oxygen originally present in the sample. The liberated iodine is titrated with standard solution of sodium thiosulphate using starch as indicator by calculating the amount of iodine liberated the DO could be determined.

Preparation of reagent -

Manganous sulphate solution - Take 250 ml titration flask weigh 91.0 gm of manganous sulphate monohydrate and dissolve it in titration flask in 9 litres of distilled water then dilute with distilled water upto 250 ml mark.

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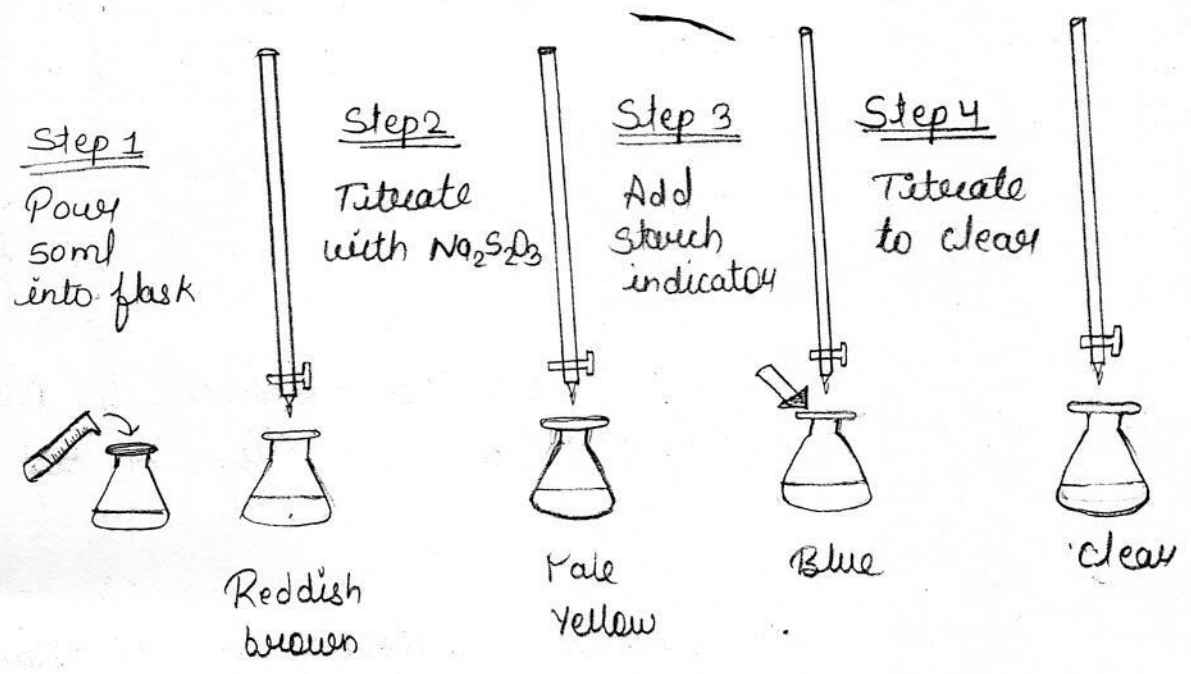


Fig: Dissolved oxygen

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ii) Alkali iodide azide reagent - In a titration flask dissolve 175 g potassium hydroxide and 37.5 g potassium iodide in a little distilled water & dilute with distilled water upto 250 ml.

Sodium thiosulphate solution - Dissolve 24.82 g Na₂S₂O₃ in a little double distilled water in a volumetric flask dilute with distilled water upto 1000 ml to make 0.025 N sodium thiosulphate solution take 250 ml of 0.1 N sodium thiosulphate & dilute it to 1000 ml with double distilled water.

Starch solution - Take 1 gm starch in a little water make a thin paste & pour it in 100 cc boiling water keep on boiling for 2 minutes and then cool.

Procedure - Collect the sample in BOD bottles add 2 ml manganous sulphate solution in the bottle by pipette. The pipette should never touch the water level it should always be above the water level.

Add 2 ml alkali iodide azide solution. Close the bottle with stopper & invert the bottle 10 times. A brown precipitate appears indicating presence of oxygen if white precipitate no oxygen.

Allow the precipitate to settle completely.

Calculation :-

Sample 1 -

$$DO_1 = \frac{V_1 \times N \times E \times 1000}{[V_4 (V_2 - V_3) / V_2]}$$

$$= \frac{1.5 \times 0.025 \times 8 \times 1000}{[50 (300 - 4) / 300]}$$

$$= \frac{300}{49.33}$$

$$= \underline{\underline{8.08 \text{ mg/l}}}$$

Sample 2

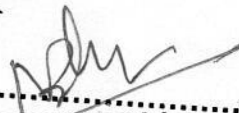
$$DO_2 = \frac{V_1 \times N \times E \times 1000}{[V_4 (V_2 - V_3) / V_2]}$$

$$= \frac{2.83 \times 0.025 \times 8 \times 1000}{[50 (300 - 4) / 300]}$$

$$= \frac{566}{(50 \times 296) / 300}$$

$$= \frac{566}{49.33}$$

$$= \underline{\underline{11.35 \text{ mg/l}}}$$


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- Remove stopper carefully & add 2ml of concentrated sulphuric acid by the side of the bottle.
- Take 203 ml of the solution from the bottle into a conical flask of 500 ml capacity.
- Titrate immediately with 0.025 N sodium thiosulphate solution using starch as indicator

Observation table -

S.No	Quantity of BOD bottle	Total volume of water sample	Burette Reading		Average reading
			Initial	Final	
1	Sample I 300 ml	50 ml	0	1	$\frac{1+1.5+2}{3}$
		50 ml	0	1.5	3
		50 ml	0	2	= 1.5 ml
2	Sample II 300 ml	50 ml	0	2.6	$\frac{2.6+2.9+3}{3}$
		50 ml	0	2.9	3
		50 ml	0	3	= 2.83 ml

Calculation -

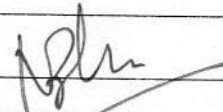
$$\text{Dissolved oxygen (mg/l)} = \frac{V_1 \times N \times E \times 1000}{[V_4 (V_2 - V_3) / V_2]}$$

- Here :
- V_1 = Volume of titrant $\text{Na}_2\text{S}_2\text{O}_3$ (ml)
 - V_2 = Total volume of water sample (ml)
 - V_3 = Volume of $(\text{MnSO}_4 + \text{KI sol}^n)$ added (ml)
 - V_4 = Volume of analyte taken in titration
 - E = Equivalent weight of O_2
 - N = Normality of $\text{Na}_2\text{S}_2\text{O}_3$ solⁿ

Result - The dissolve oxygen of water sample 1 = 6.08 mg/l

The dissolve oxygen of water sample 2 = 11.35 mg/l

Ayasha


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Date sheet - Sample Ist

Sample : Out side plant nursery Kalwar, Jaipur

Date of sample collection - 7 february 2024

Time : 11 Am

Amount : 300ml + 300ml

Day 1 : 7 february 2024

Day 5 : 12 february 2024

Date sheet - Sample IInd

Sample : Tank farm farm


Date of sample collection : 15 february 20.

Time : 9 Am

Amount : 300ml + 300ml

Day 1 : 15 february 2024

Day 5 : 19 february 2024


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Experiment - 14

Aim:

Determination of BOD in water sample.

Requirements - Titration flask, titration tube, manganous sulphate solution, Alkali iodide azide reagent, Concentrated sulphuric acid, sodium thiosulphate solution & starch solution.

Principle - Biochemical oxygen demand is the amount of oxygen taken by the micro-organism that decompose the organic waste matter.

→ BOD correlate with organic matter present in water body / sample which in term is a measure of pollution level.

→ The test has its widest application in measuring waste loading to treatment plant and is evaluating the efficiency of such treatment system.

→ It is calculated by estimating dissolved oxygen at two different time points.

Preparation of reagent -

(i) Manganous sulphate solution - Take 250 ml titration flask weigh 91.0 gm of manganous sulphate monohydrate and dissolve it in titration flask in 9 litre of distilled water. Then dilute with distilled water upto 250 ml mark.

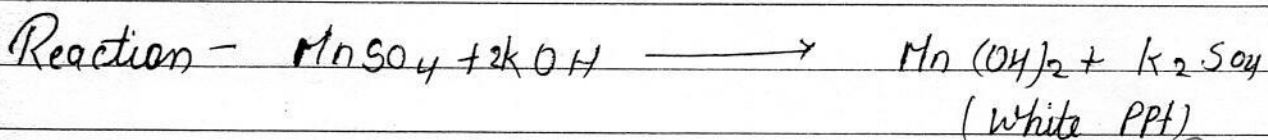
(ii) Alkali iodide oxide reagent - In a titration flask dissolve 175 g potassium hydroxide and 37.5 g potassium iodide in a little distilled water & dilute with distilled water upto 250 ml.

(iii) Sodium thiosulphate solution - Dissolve 24.82 g sodium thiosulphate in a little double distilled water in a volumetric flask dilute with distilled water upto 1000 ml to make 0.025 N sodium thiosulphate & dilute it to 1000 ml with double distilled water.

(iv) Starch solution - Take 1 gm starch in a little water make a thin paste & pour it in 100 cc boiling water keep on boiling for 2 minutes & then cool.

Procedure - Collect water sample in 2 BOD bottle without making air bubbles.

- In one bottle calculate dissolved oxygen that is DO_1
- Another BOD bottle place in BOD incubator for 5 days where temperature is almost constant i.e. $20^\circ C$
- After 5 days calculate the DO of 2nd BOD bottle that is DO_2
- Difference between these two DO that is DO_1 & DO_2 is BOD



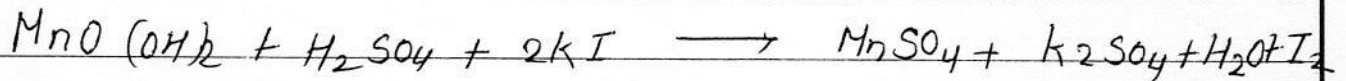
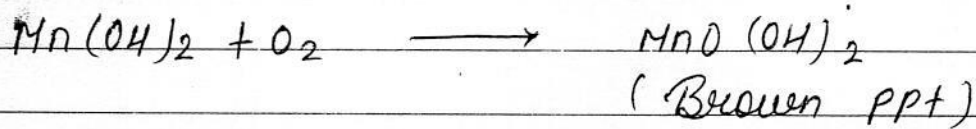
Calculation

Sample 1

$$\begin{aligned} DO_1 &= \frac{V_3 \times N \times 8 \times 1000}{V_2 \left(\frac{V_1 - V_4}{V_1} \right)} \\ &= \frac{2.83 \times 0.025 \times 8 \times 1000}{50(300-4) / 300} \\ &= \frac{566}{49.33} \\ &= \underline{\underline{11.47}} \text{ mg/L} \end{aligned}$$

$$\begin{aligned} DO_5 &= \frac{V_3 \times N \times 8 \times 1000}{V_2 (V_1 - V_4) / V_1} \\ &= \frac{1.5 \times 0.025 \times 8 \times 1000}{50(300-4) / 300} \\ &= \frac{300}{49.33} \\ &= \underline{\underline{6.081}} \text{ mg/L} \end{aligned}$$

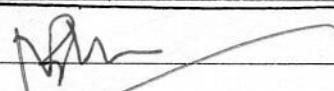
$$\begin{aligned} BOD &= DO_1 - DO_5 \\ &= 11.47 - 6.081 \\ &= \underline{\underline{5.389}} \text{ mg/L} \end{aligned}$$



Observation table :-

S.No	Quantity of BOD bottle	Total volume of water sample	Burette reading		Average reading
			Initial	Final	
1	Sample 1 st DO ₁ 300 ml	50 ml	0	2.9 ml	$\frac{2.9 + 3.0 + 2.6}{3} = 2.83 \text{ ml}$
		50 ml	0	3.0 ml	
		50 ml	0	2.6 ml	
2	(DO ₂)	50 ml	0	1 ml	$\frac{1 + 2 + 1.5}{3} = 1.5 \text{ ml}$
		50 ml	0	2 ml	
		50 ml	0	1.5 ml	

S.No	Quantity of BOD bottle	Total volume of water sample	Burette reading		Average reading
			initial	final	
1	Sample 2 (DO ₁) 300 ml	50 ml	0	4.5	$\frac{4.5 + 4.9 + 4.7}{3} = 4.7$
		50 ml	0	4.9	
		50 ml	0	4.7	
2	(DO ₂) 300 ml	50 ml	0	1.3	$\frac{1.3 + 1.2 + 1.2}{3} = 1.23$
		50 ml	0	1.2	
		50 ml	0	1.2	


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Calculation

Sample 2

$$\begin{aligned} DO_1 &= \frac{V_3 \times N \times 8 \times 1000}{V_2 (V_1 - V_2) / V_1} \\ &= \frac{4.7 \times 0.025 \times 8 \times 1000}{50(300-4) / 300} \\ &= \frac{940}{49.33} \\ &= \underline{\underline{19.05 \text{ mg/L}}} \end{aligned}$$

$$\begin{aligned} DO_5 &= \frac{V_3 \times N \times 8 \times 1000}{V_2 (V_1 - V_4) / V_1} \\ &= \frac{1.23 \times 0.025 \times 8 \times 1000}{50(300-4) / 300} \\ &= \frac{246}{49.33} \\ &= \underline{\underline{4.98 \text{ mg/L}}} \end{aligned}$$

$$\begin{aligned} BOD &= DO_1 - DO_5 \\ &= 19.05 - 4.98 \\ &= \underline{\underline{14.07 \text{ mg/L}}} \end{aligned}$$


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Calculation -

$$\text{formula} - \frac{V_3 \times N \times 8 \times 1000}{V_2 (V_1 - V_4)}$$

where V_1 = Volume of BOD bottle = 300ml

V_2 = Sample used = 50ml

V_3 = titration volume of sodium thiosulphate

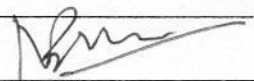
V_4 = Volume of $MnSO_4$ + alkali azide = 4ml

Result -

1. Biological O_2 demand (BOD) of sample-1
= 5.389 mg/L

2. Biological O_2 demand (BOD) of sample-2
= 14.07 mg/L

Ans
21/12/24



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